

# DNA supercoiling: plectonemes or curls ?

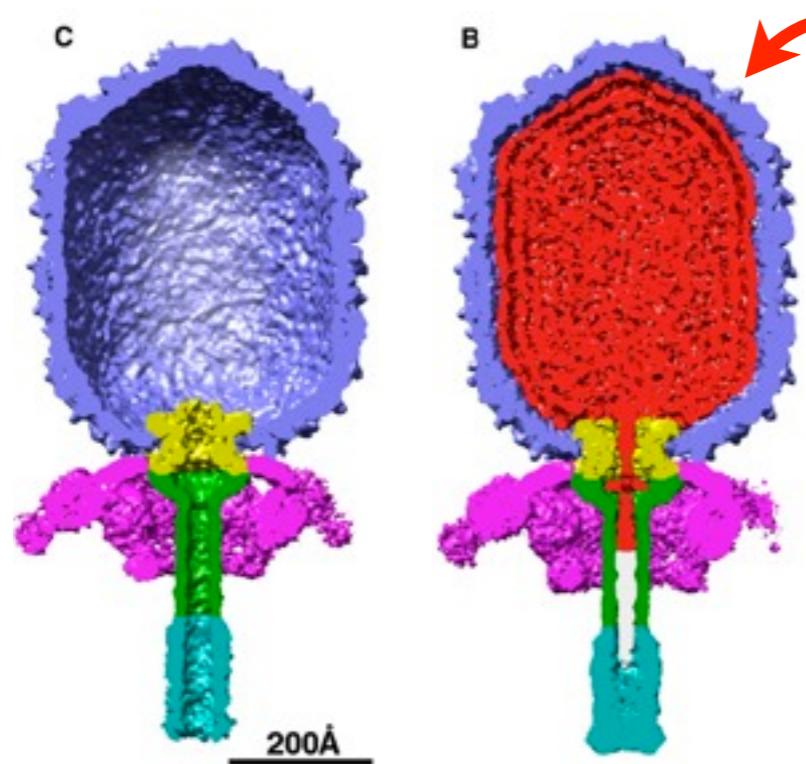
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Northwestern University (IL. USA)

# Why study DNA mechanical properties ?

mechanical properties influence biology of the cell

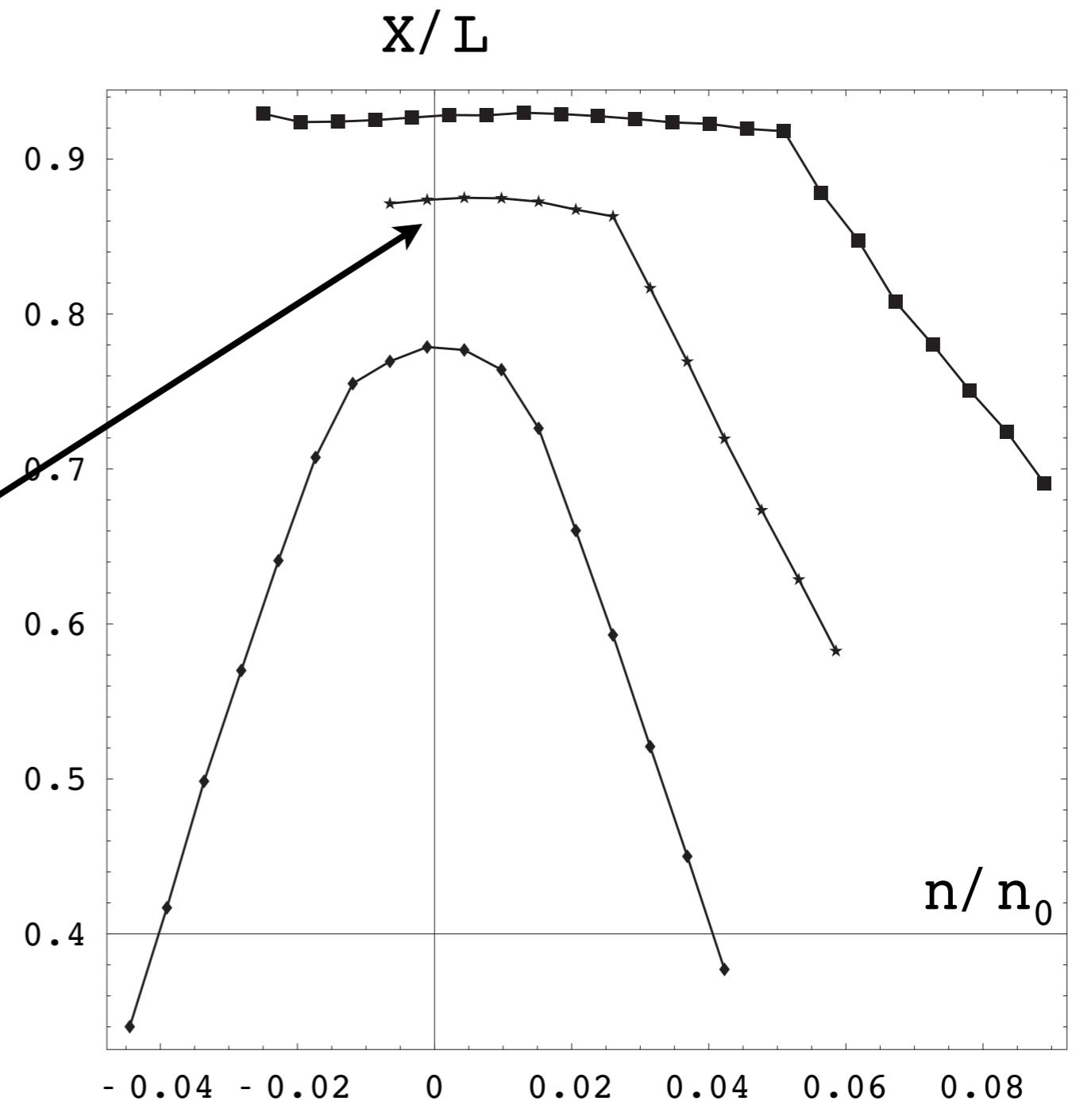
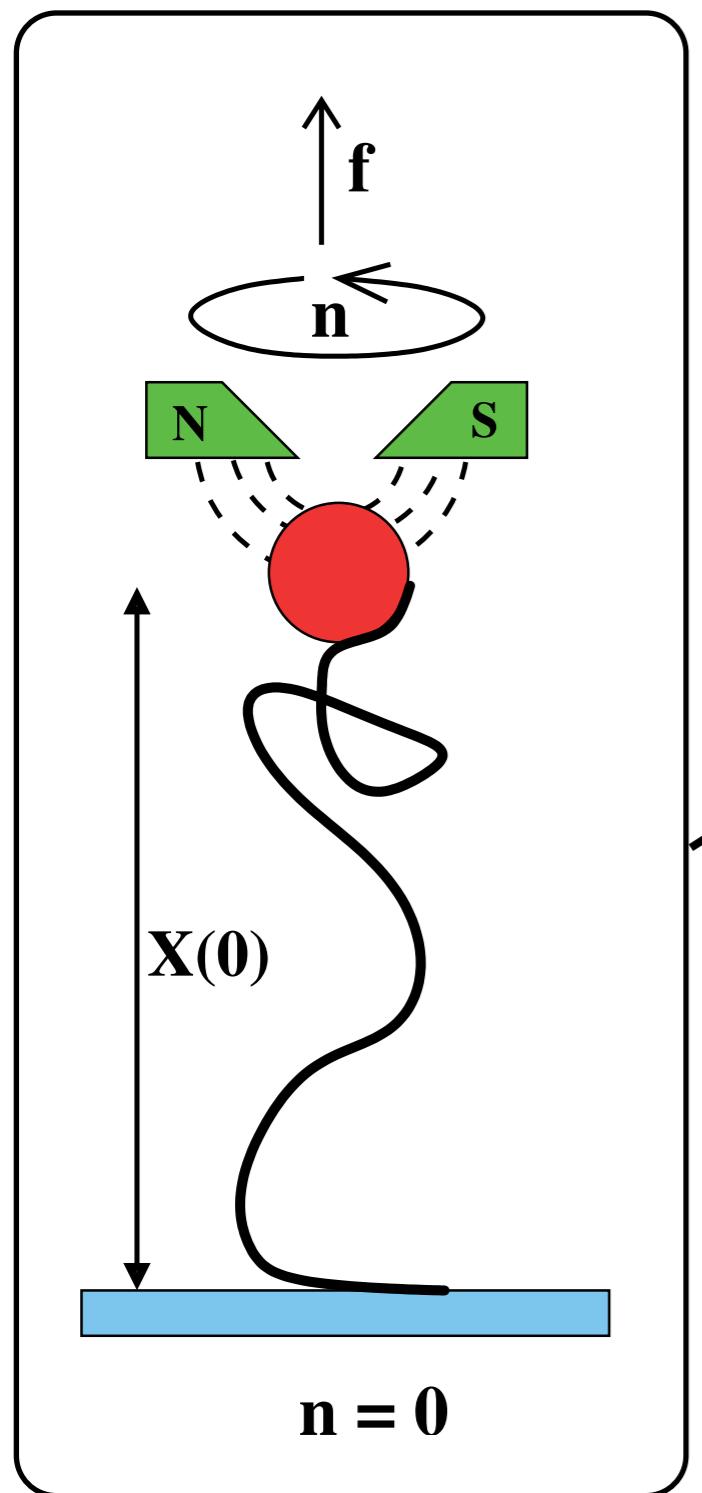
- 1 meter of DNA in a 10 micron wide nucleus
- ejection from viral capsid
- transcription (RNAPolymerase is torque dependent)
- protein binding is strain dependent, or induces strain on DNA
- chromatin compaction/decompaction (cell division)



20kbp dsDNA (6800nm)  
in 40nm wide capsid

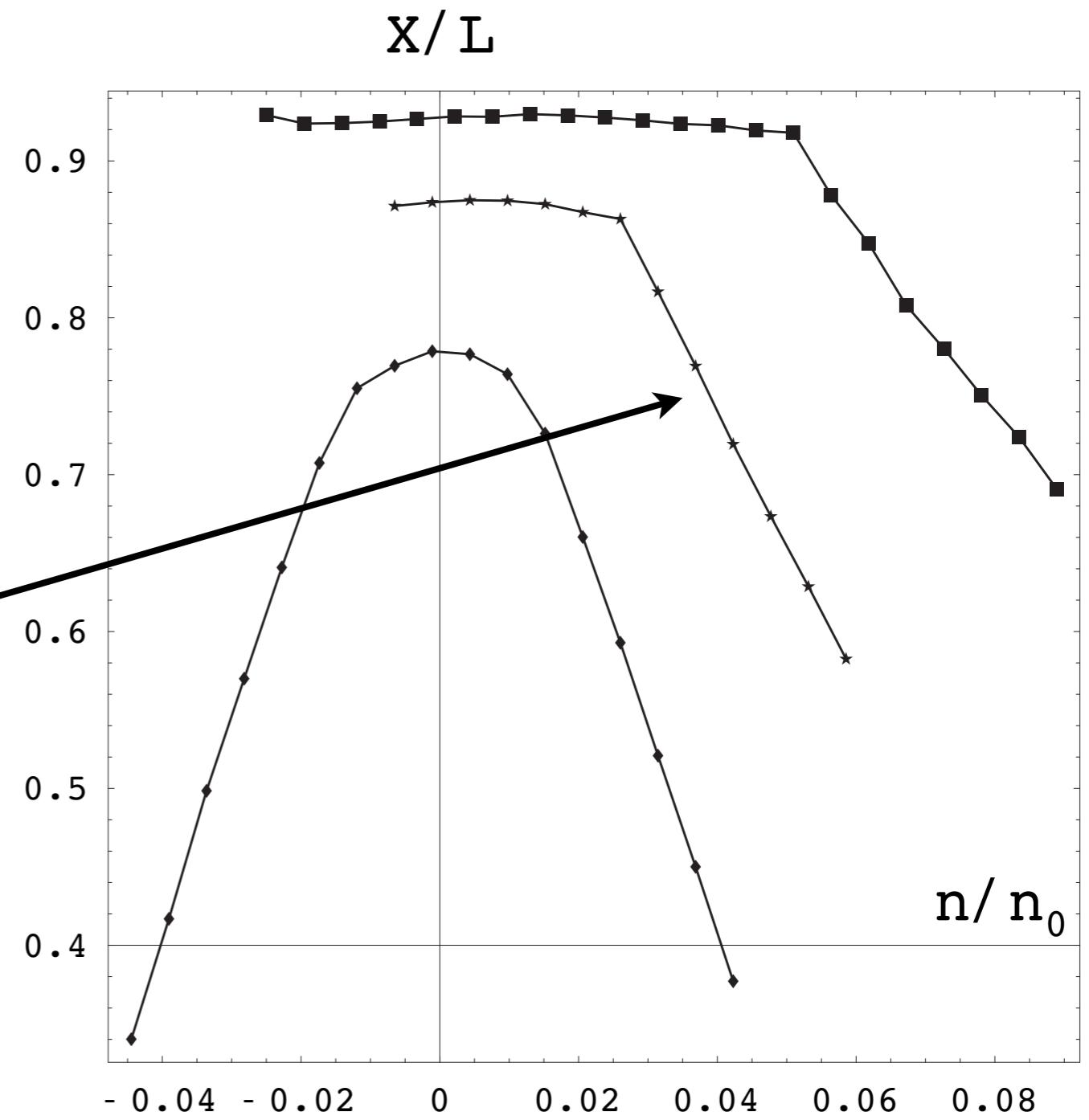
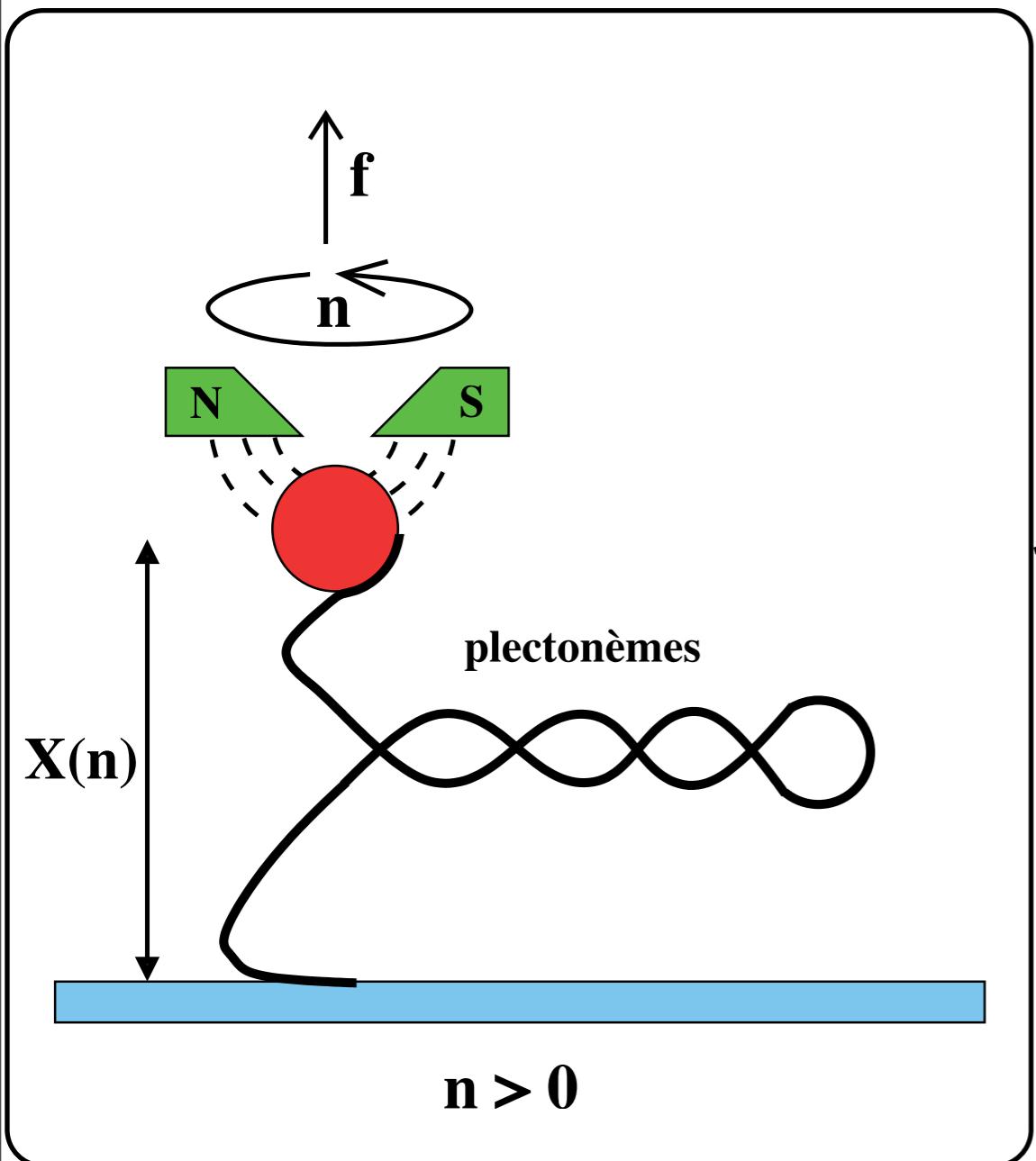
Tang et al (Structure) 2008

# Pulling and twisting DNA



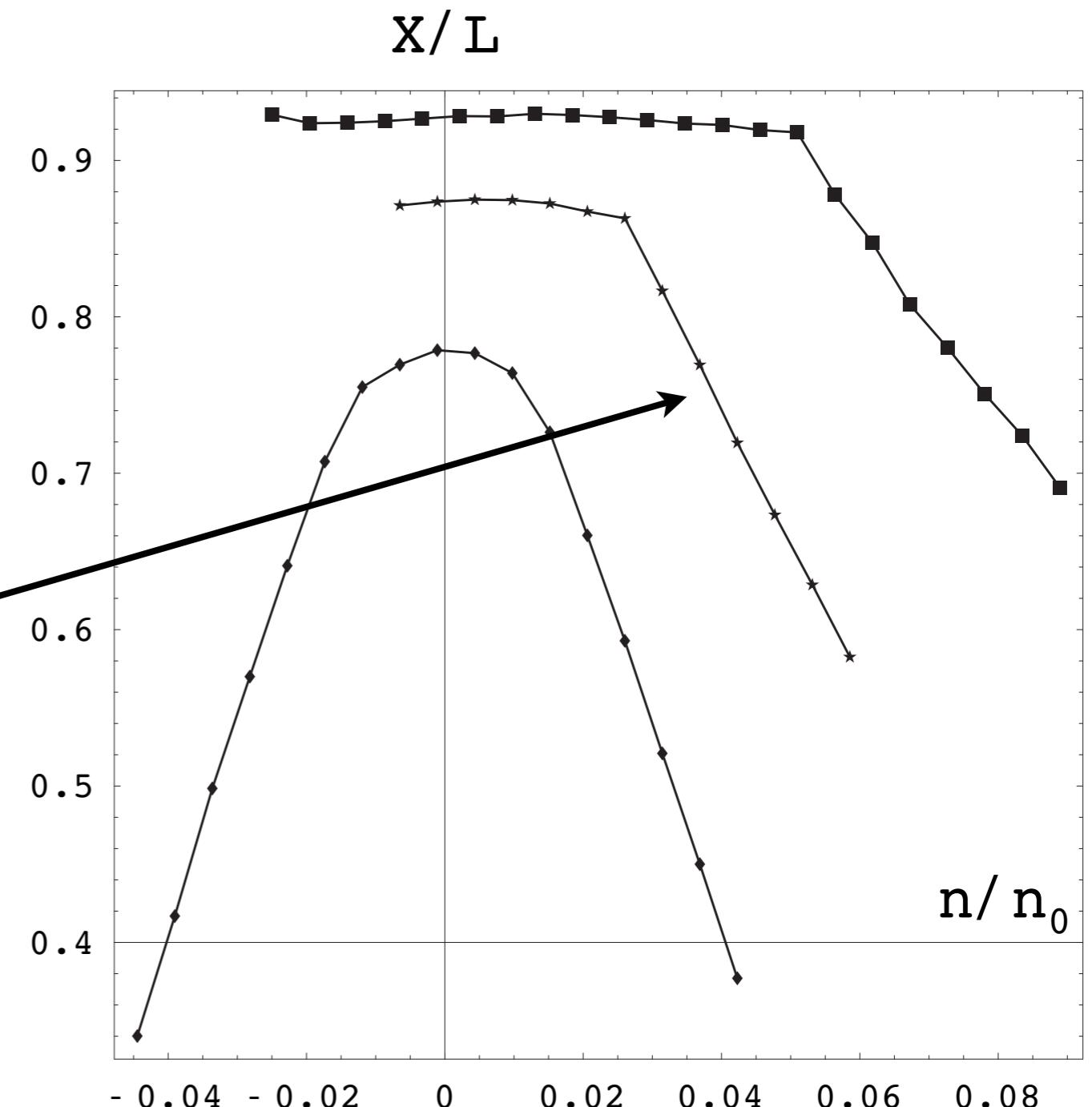
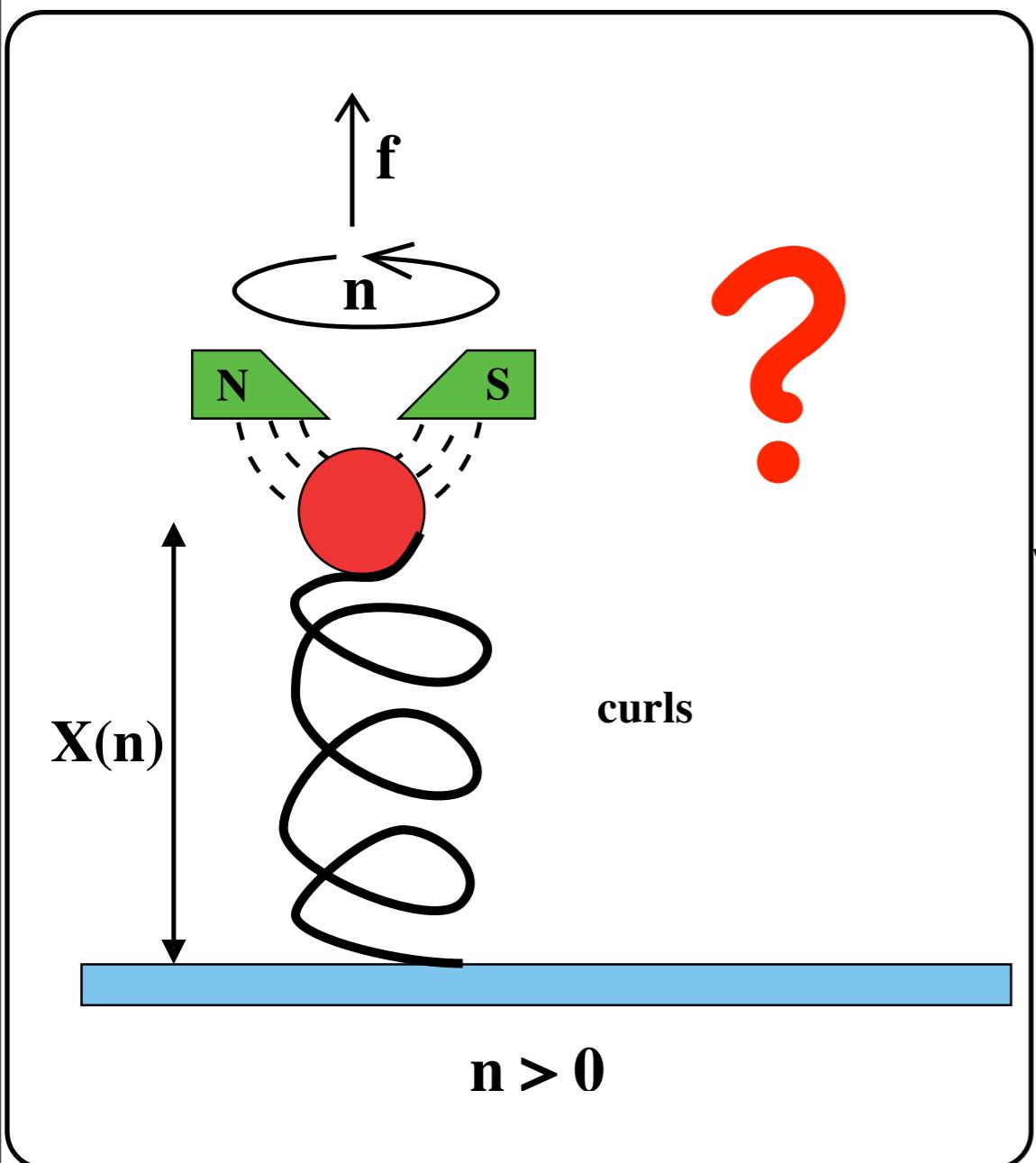
Data from G. Charvin (LPS-ENS)

# Pulling and twisting DNA



Data from G. Charvin (LPS-ENS)

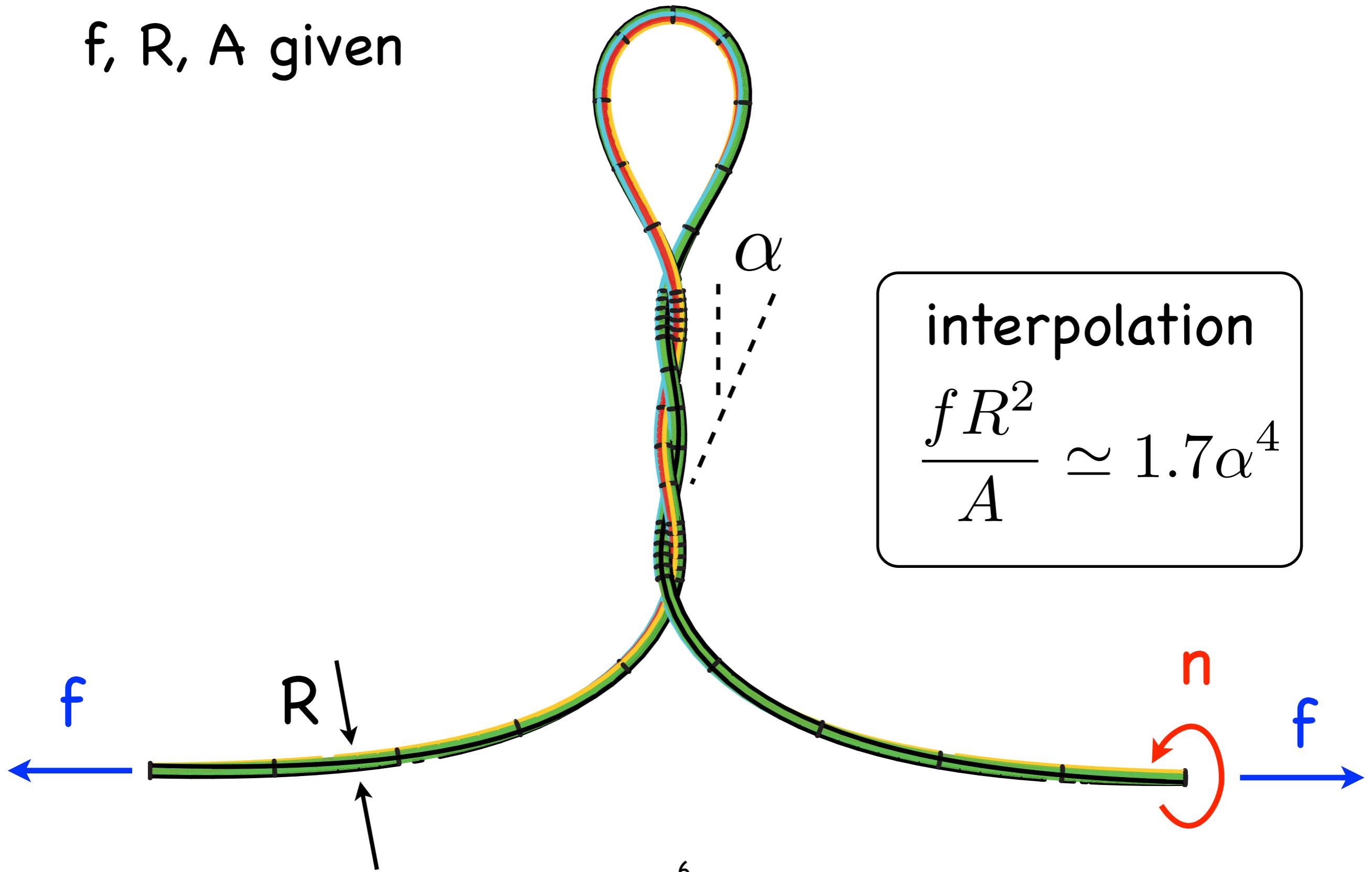
# Pulling and twisting DNA



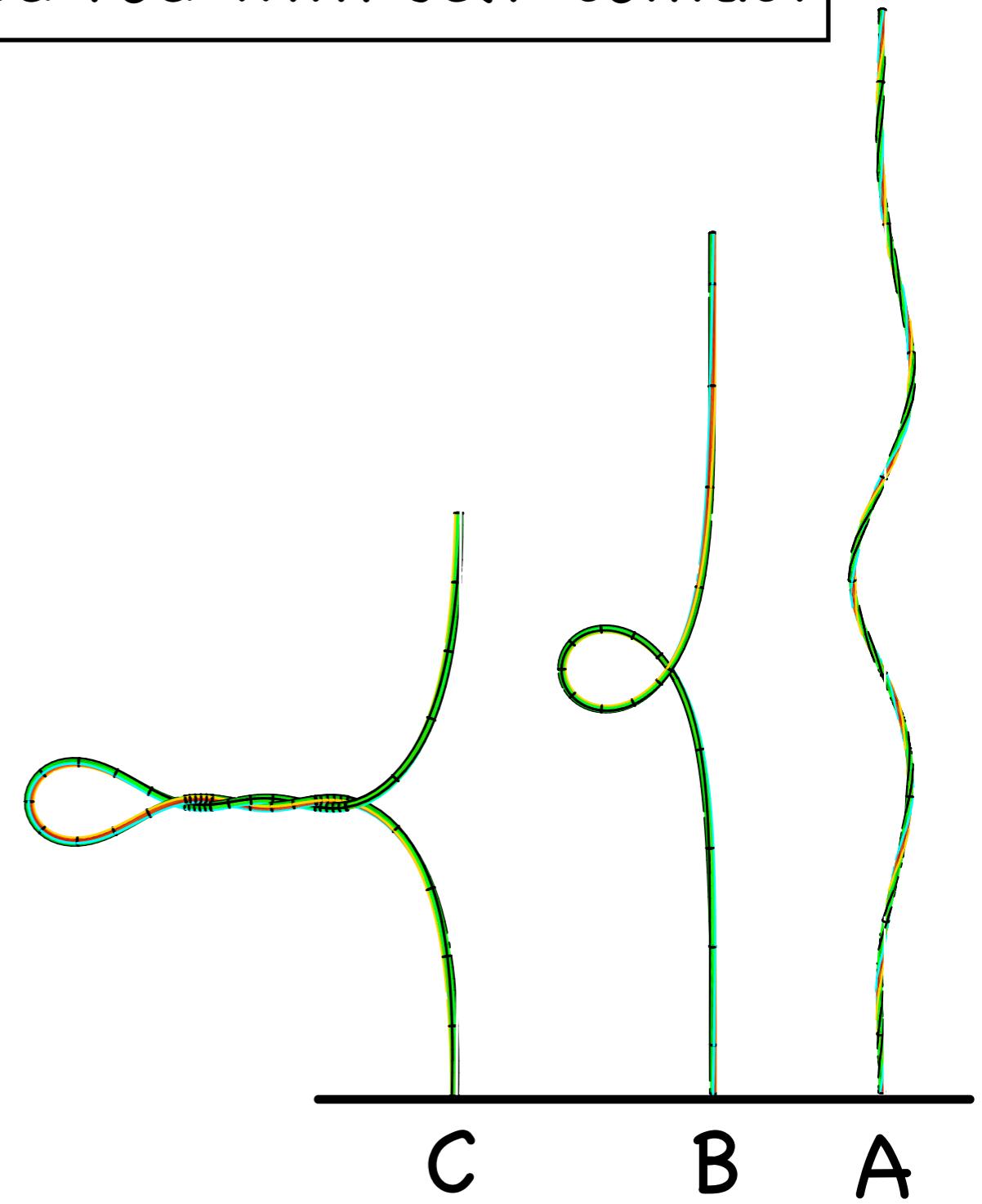
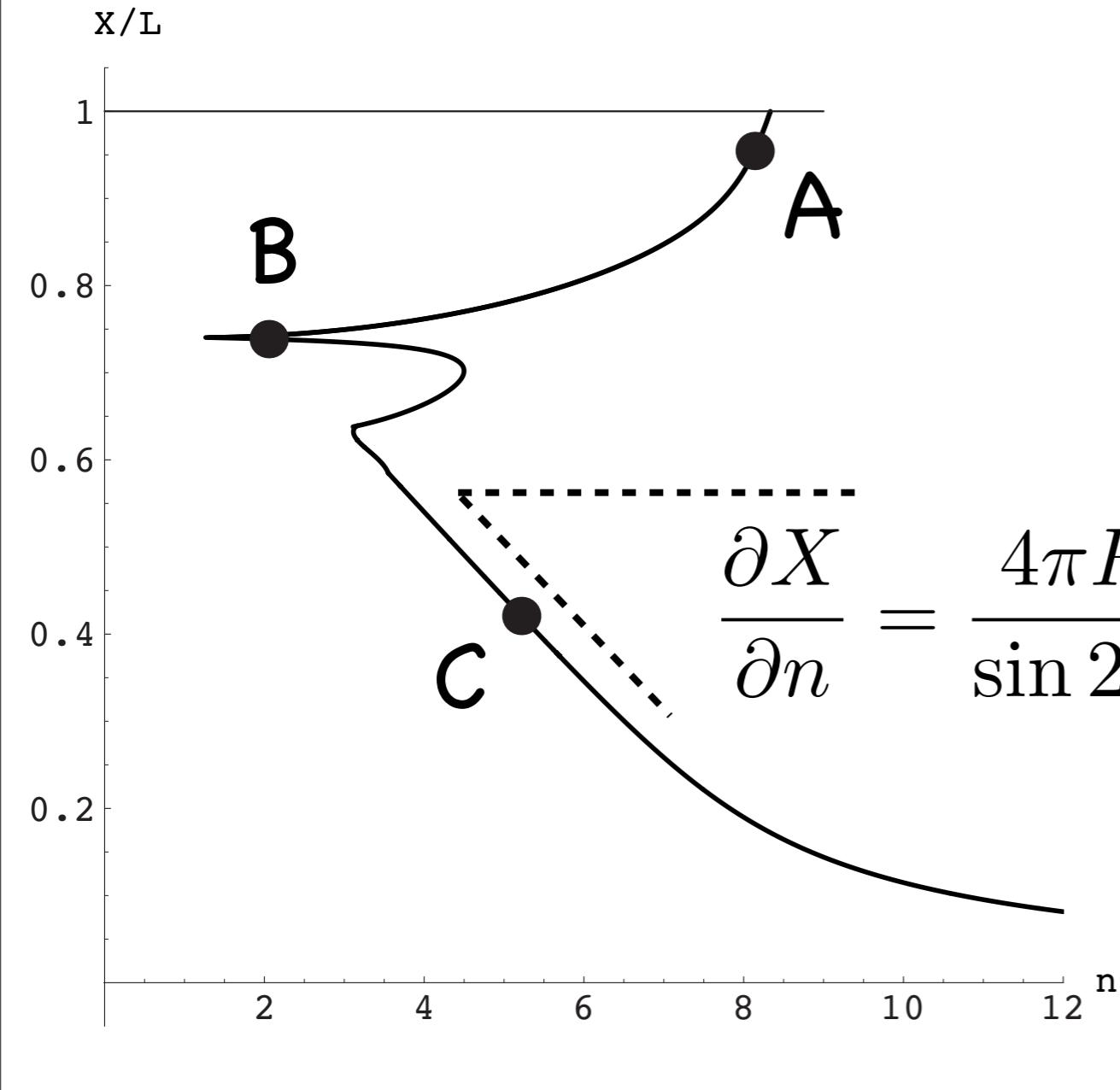
Data from G. Charvin (LPS-ENS)

# Numerical simulations : twisted rod with self-contact

$f, R, A$  given



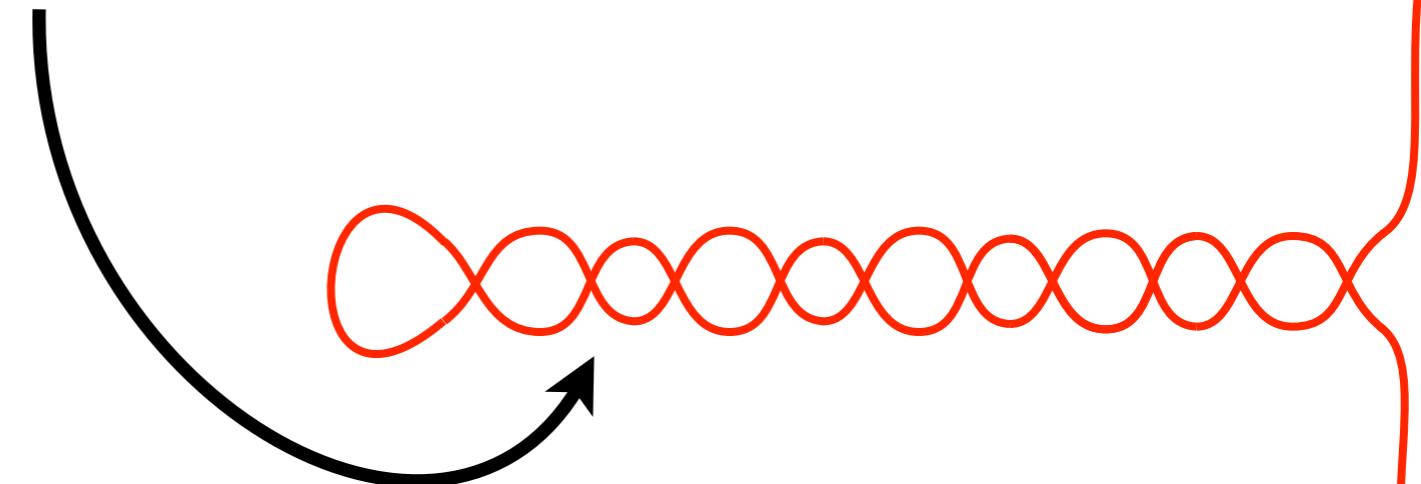
# Numerical simulations : twisted rod with self-contact



(based on Swigon+Coleman model for contact in Kirchhoff rods)

## Limitations of the twisted rod model

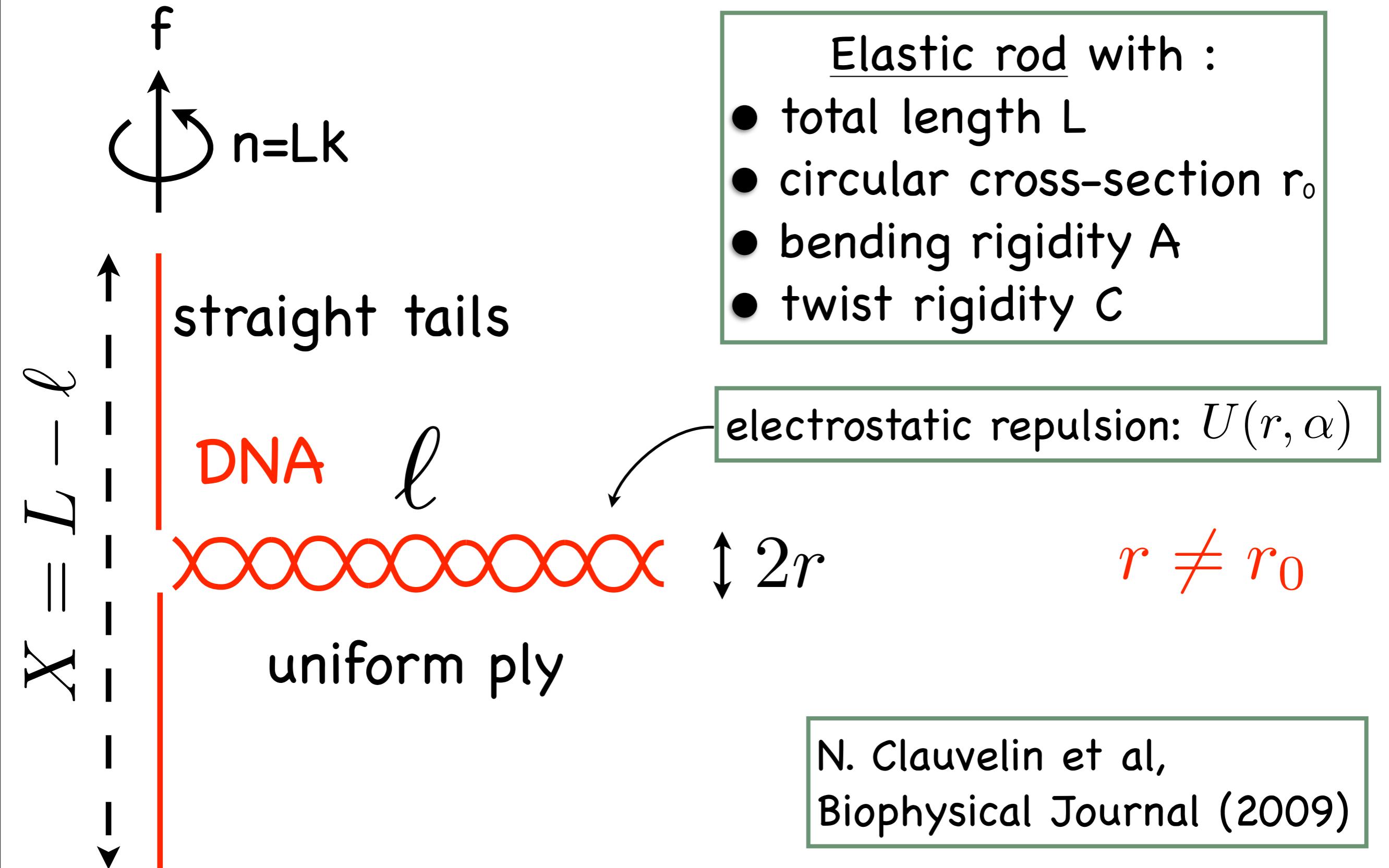
1. Electrostatics repulsion :  
supercoiling radius  $R$  **is not** 1 nm



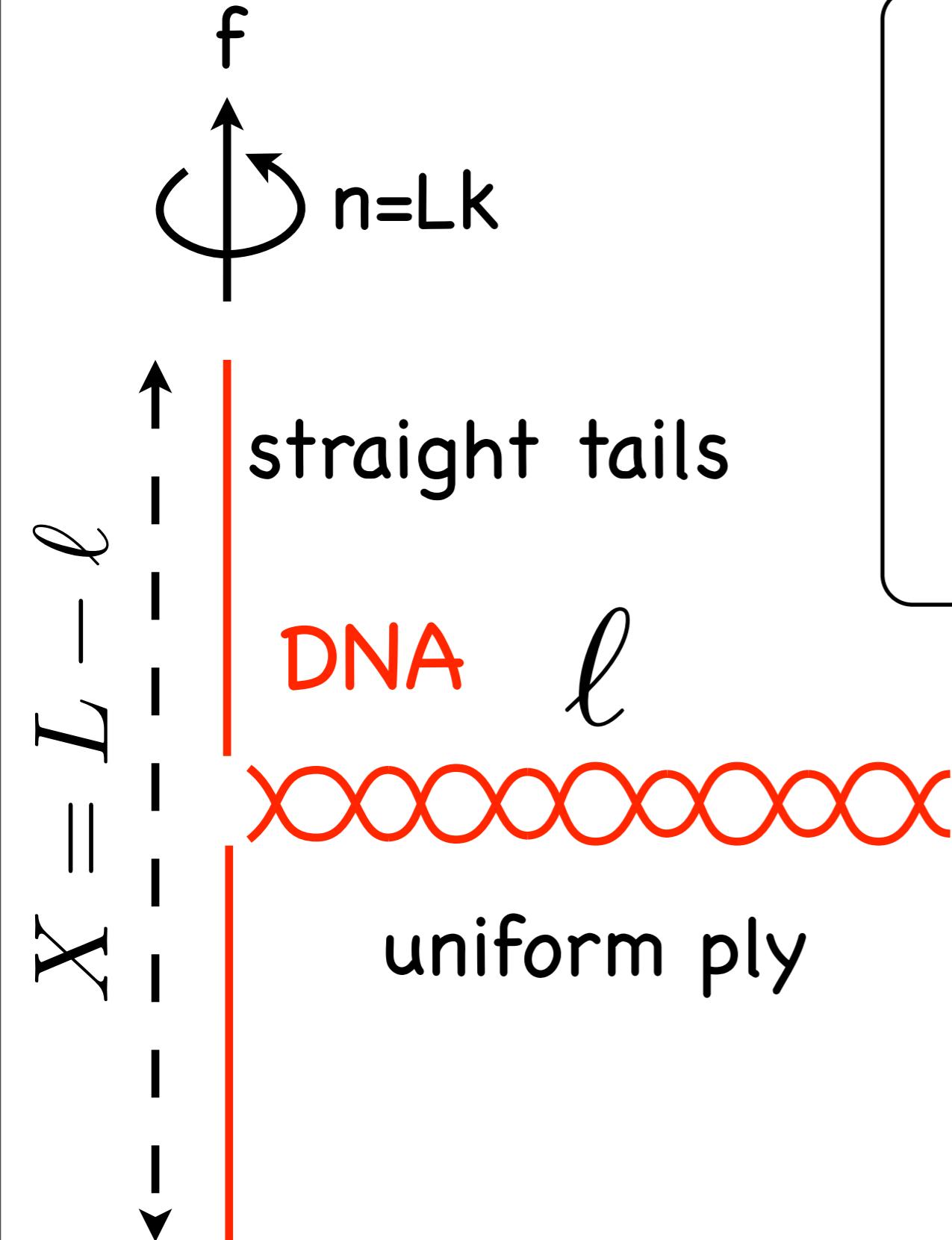
2. Tails are not straight :  
«disorded walk» (Worm Like Chain)



# Analytical model with electrostatics repulsion



# Analytical model with electrostatics repulsion



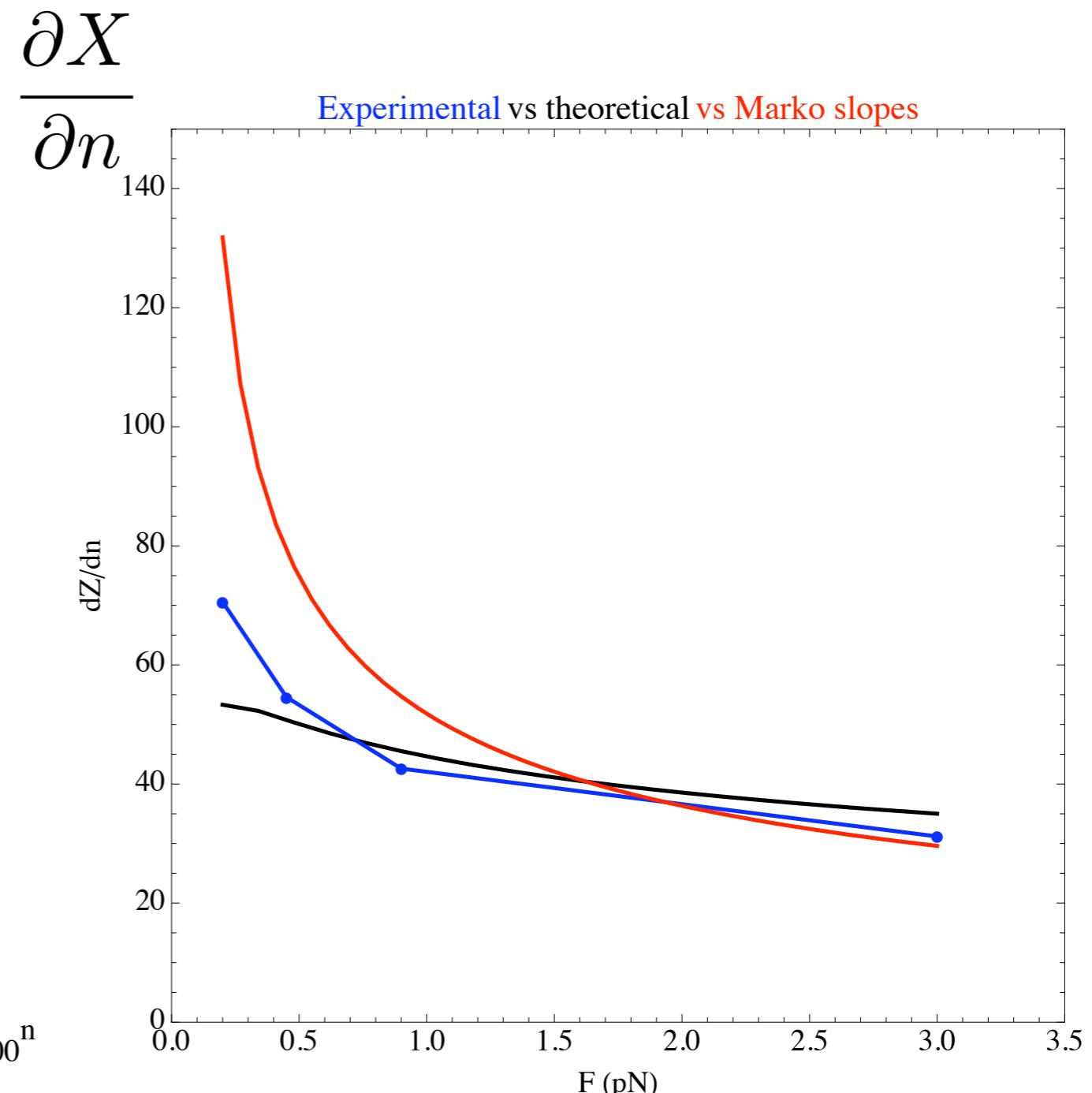
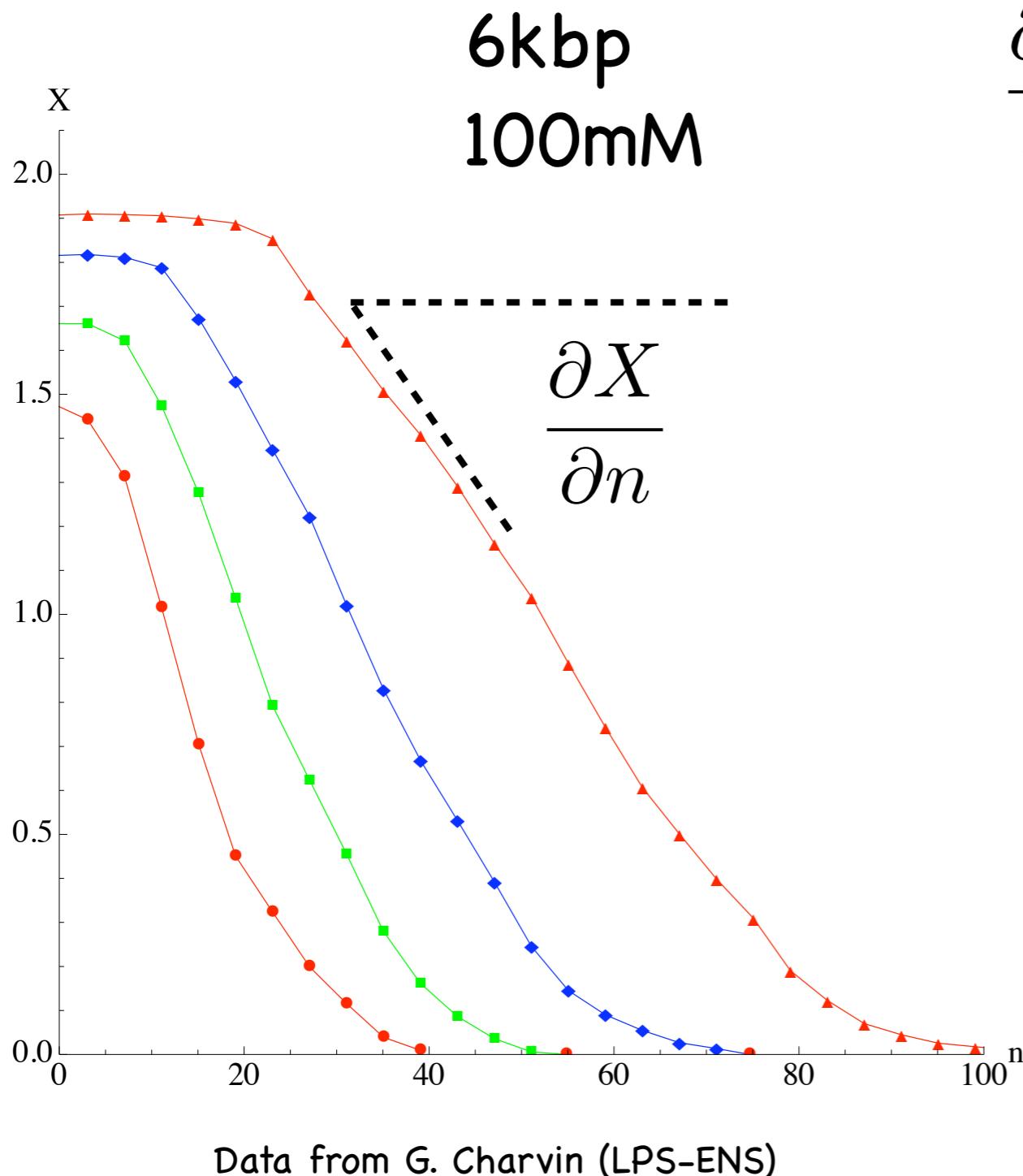
$$V = \frac{1}{2}A \frac{\sin^4 \alpha}{r^2} \ell \quad \text{bending}$$
$$+ \frac{1}{2}C\tau^2 L \quad \text{twisting}$$
$$+ U(R, \alpha)\ell \quad \text{electro}$$
$$- fX \quad \text{ext. force}$$

with constraint:

$$n = Lk = \frac{1}{2\pi} \left( \tau L + \frac{\sin 2\alpha}{2r} \ell \right)$$

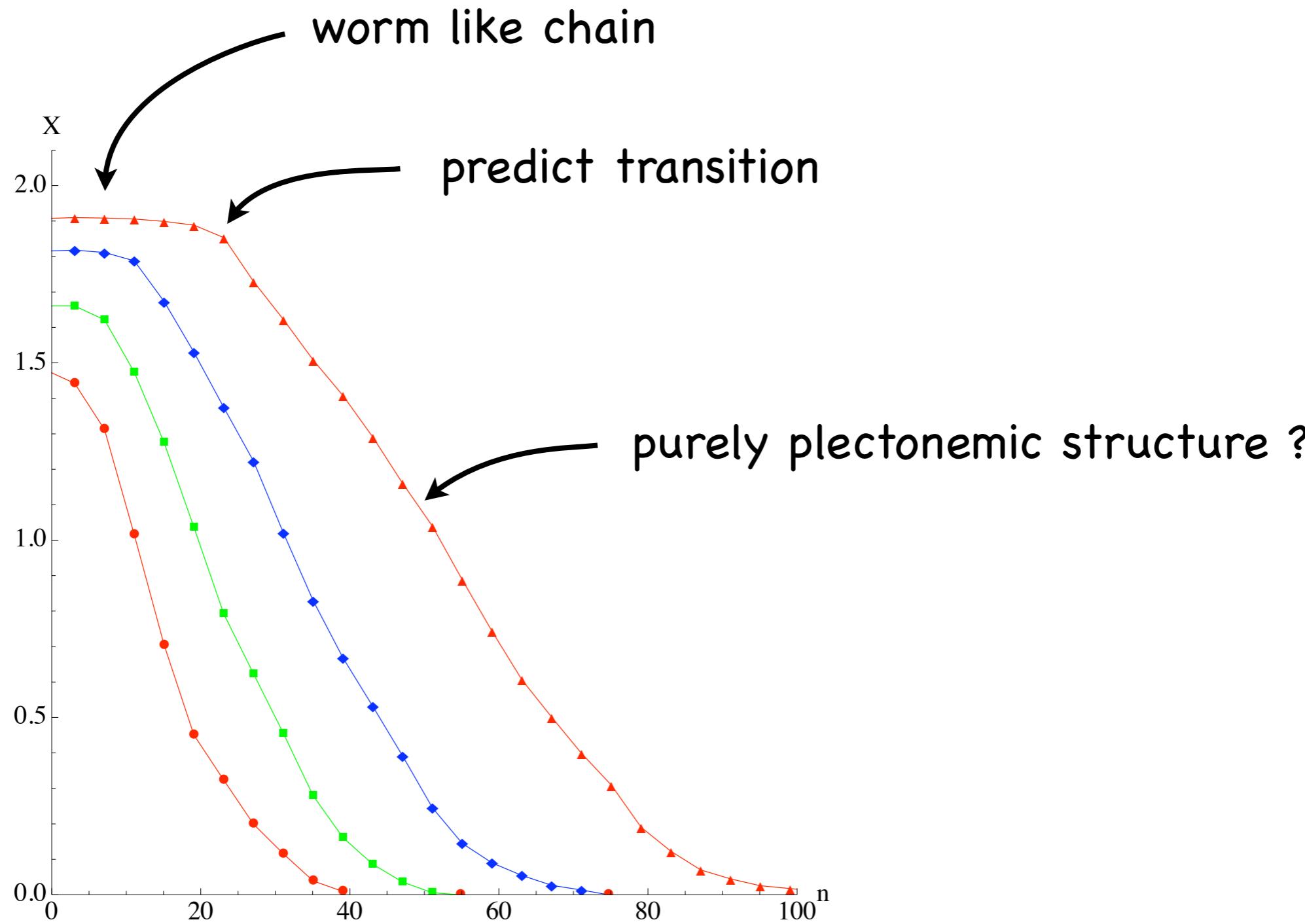
minimize  $V \Rightarrow (\alpha, r, \tau, \ell)$

# Analytical model with electrostatics repulsion : results

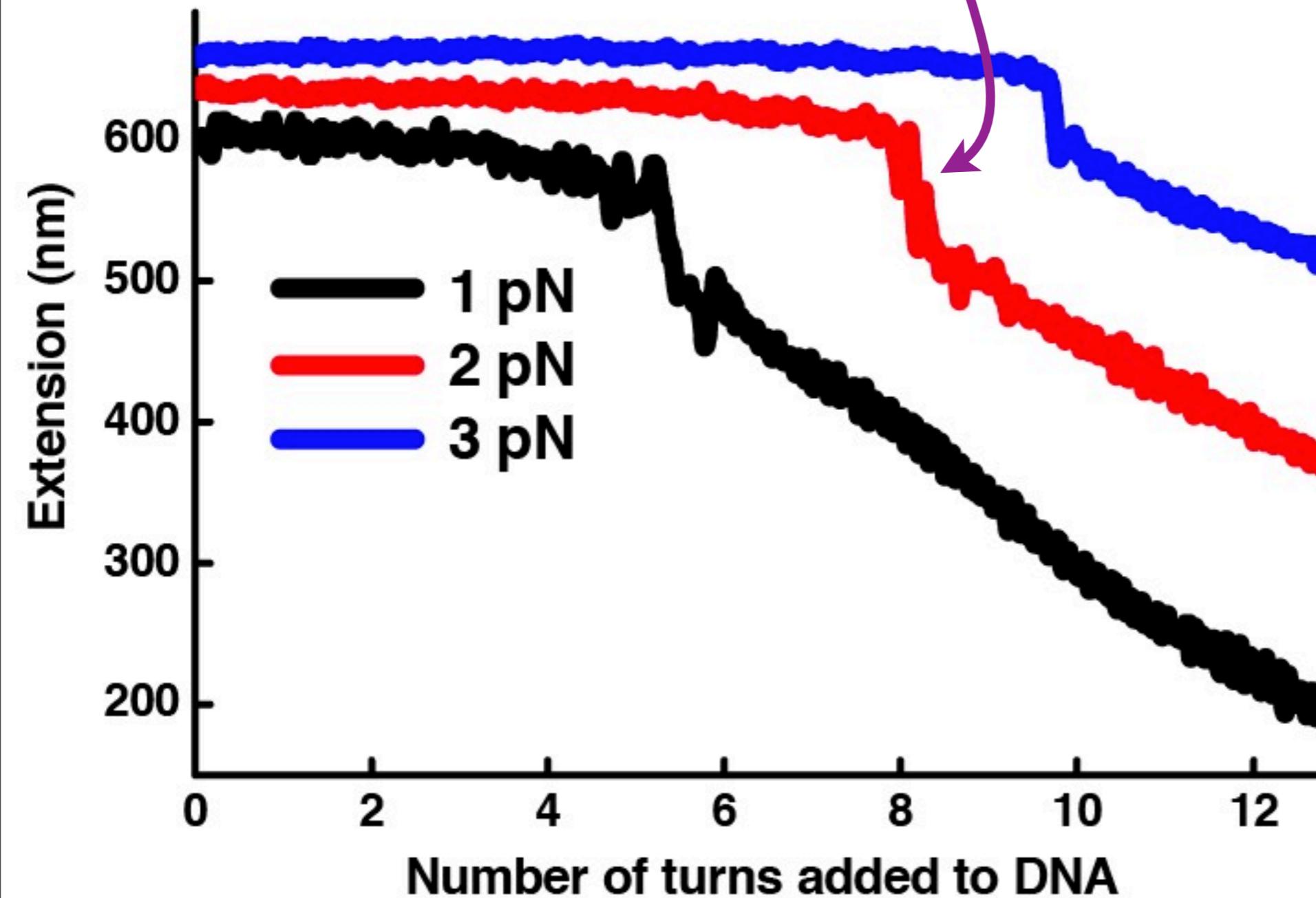


J. Marko, "Torque and dynamics of linking number ...", Phys. Rev. E (2007)

# Statistical mechanics model



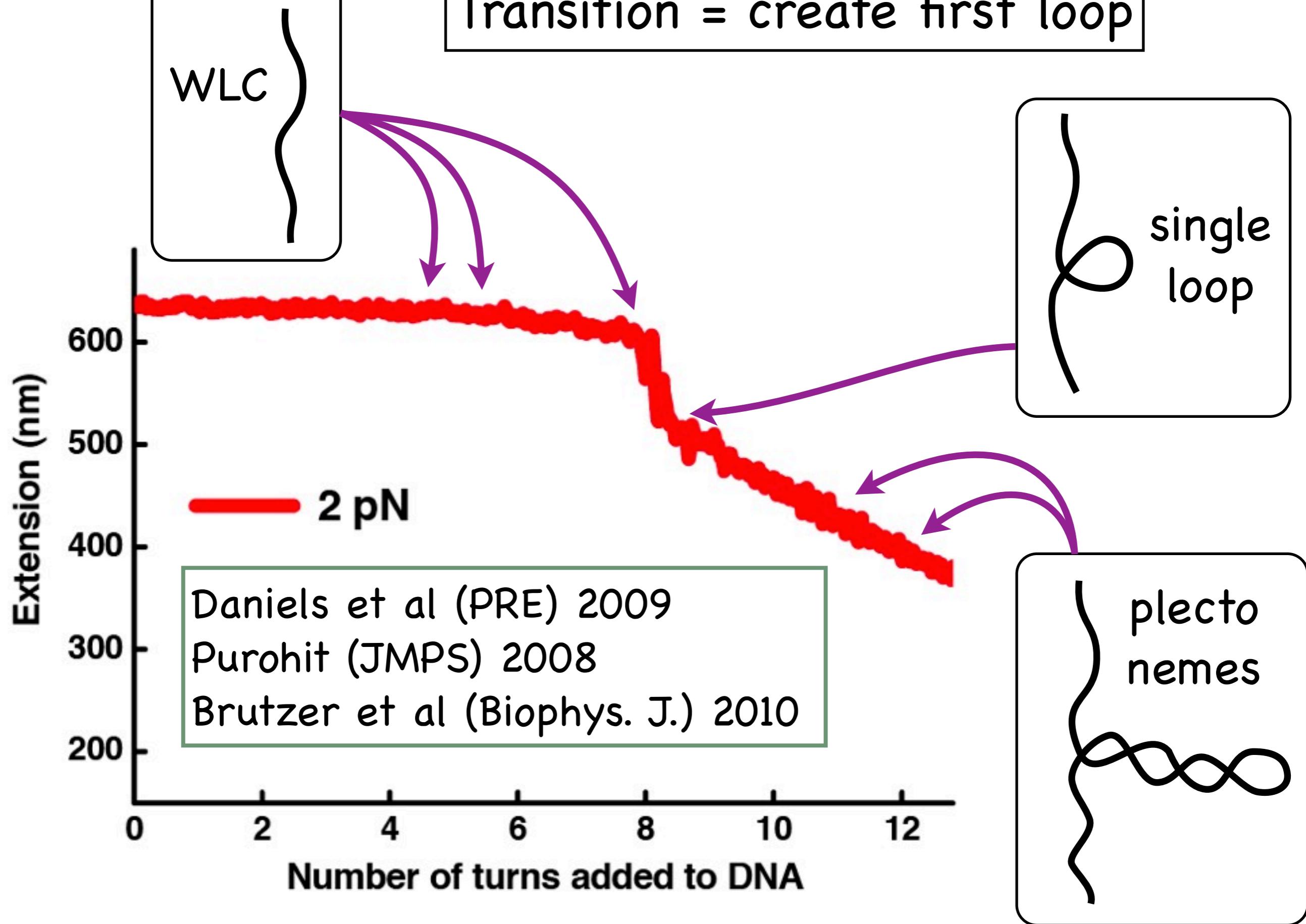
# Abrupt transition



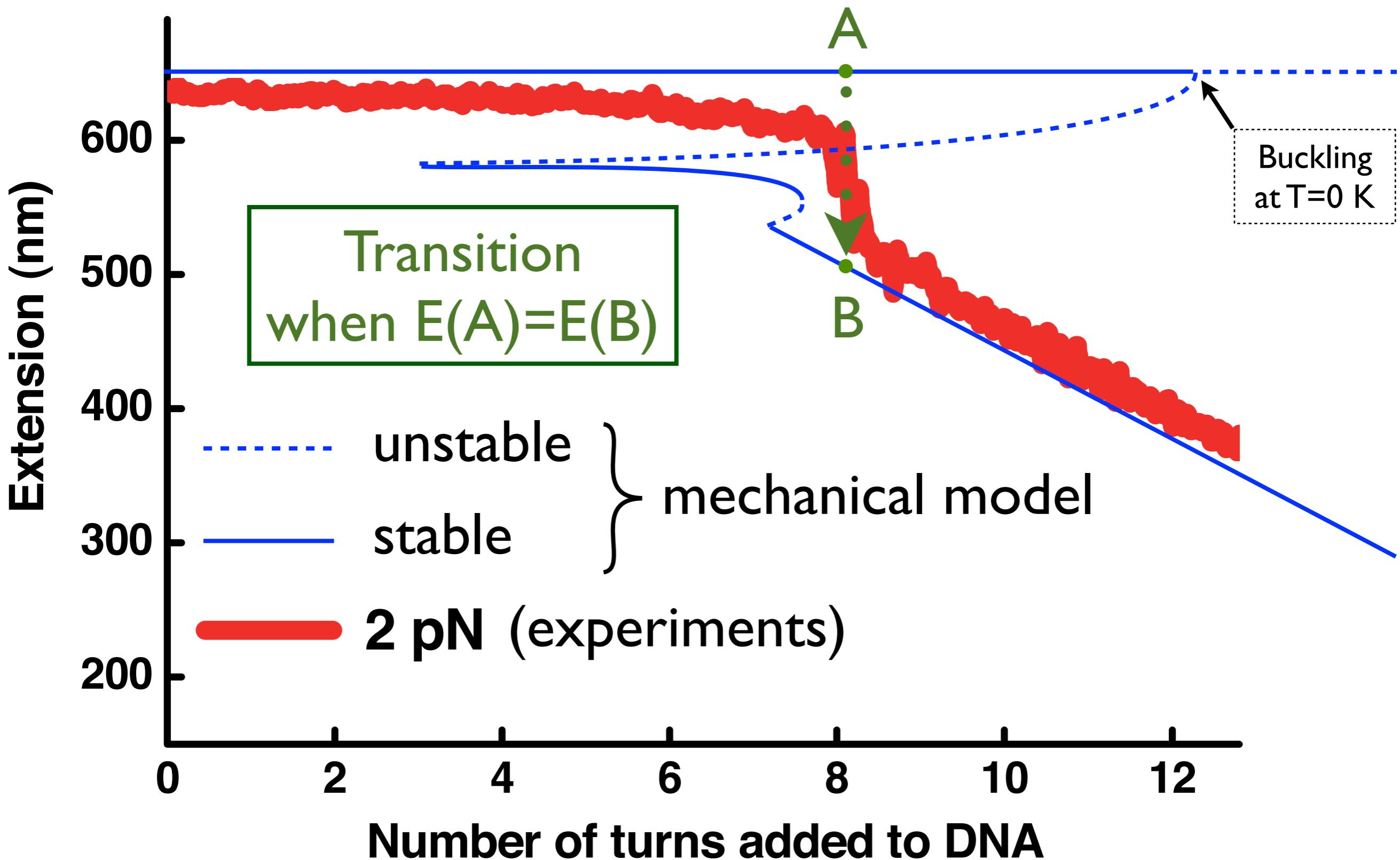
2.2 kbp  
150 mM

Forth et al  
Phys. Rev. Lett.  
2008

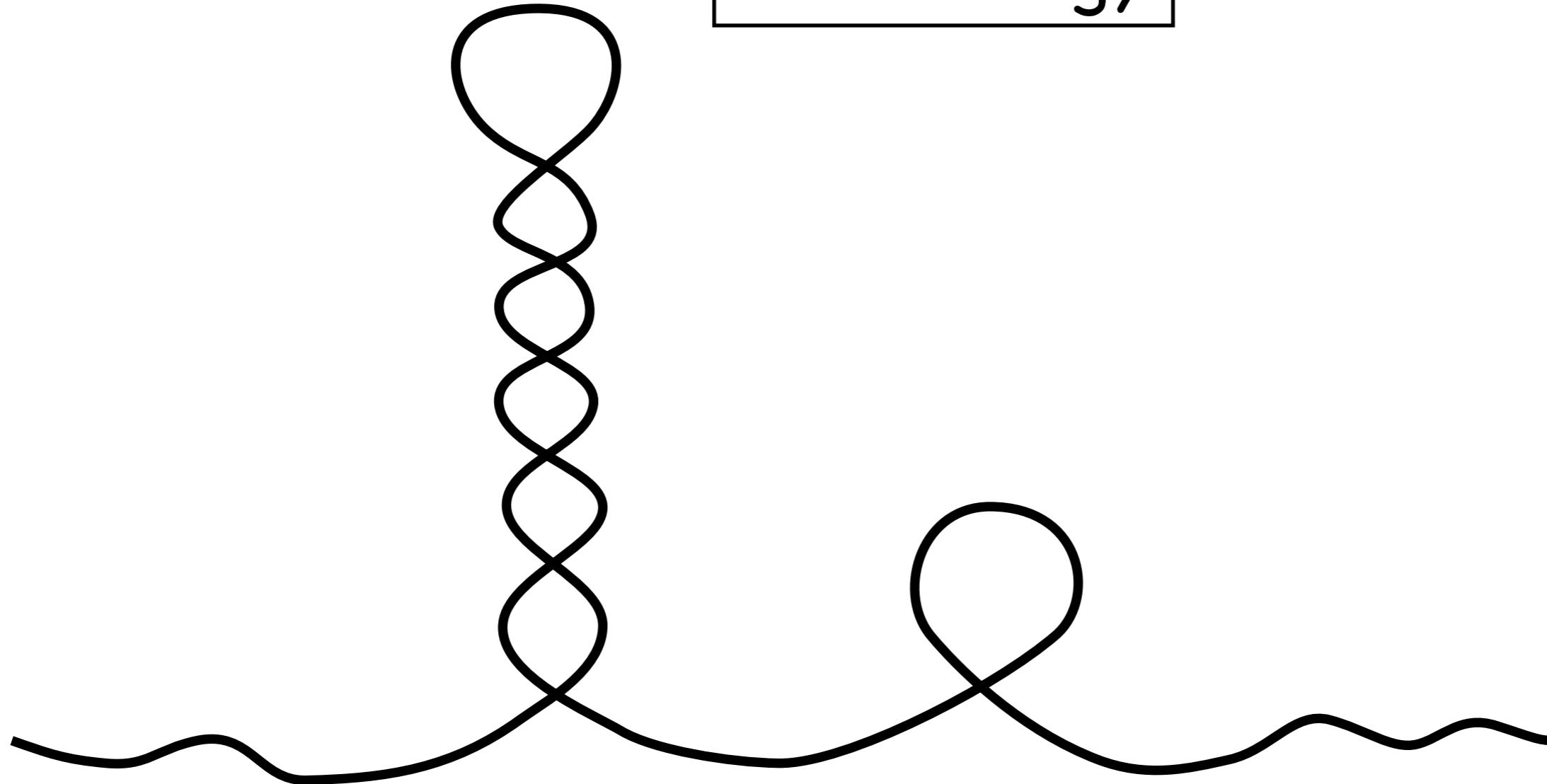
Transition = create first loop



# Comparing free-energies of straight and supercoiled DNA



Free energy



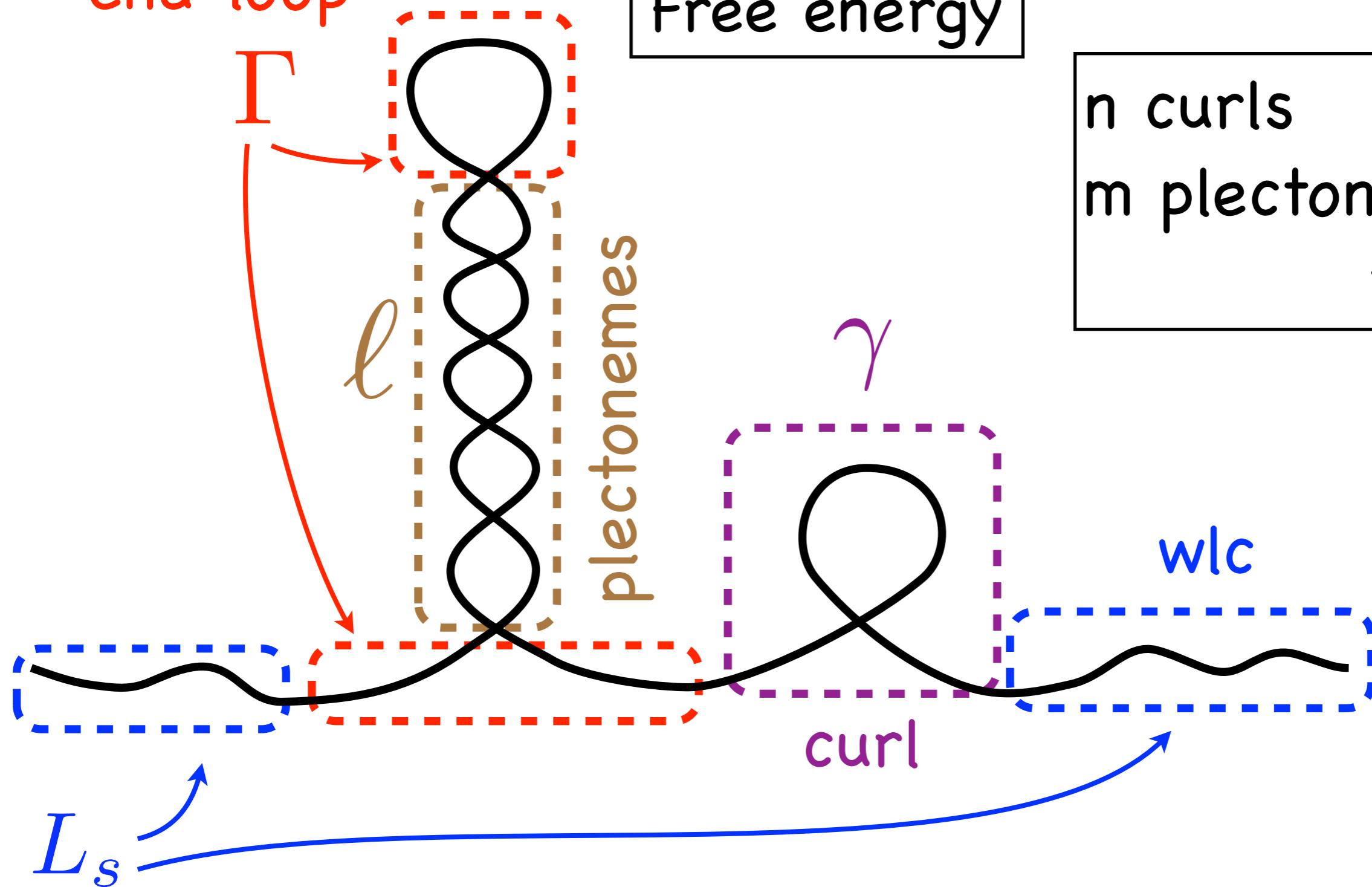
total contour length

$$L = \text{nbp} \cdot 0.34 \text{ nm}$$

end-loop

$\Gamma$

Free energy



n curls

m plectonemic  
regions

$$L = \text{nbp} \cdot 0.34 \text{ nm} = L_s + n\gamma + m(\Gamma + \ell)$$

# Free energy

corrections to the work of the external force

$$L = L_s + n\gamma + m(\ell + \Gamma)$$

$$F = F_s + n F_\gamma + m (F_\Gamma + F_\ell) + F_f - T S$$


---

entropic terms

$$F_s = E_{twist} + E_{bend} + E_f = \frac{1}{2} C_s \tau_s^2 L_s - g(f) L_s$$

$$F_\gamma = E_{twist} + E_{bend} = \frac{1}{2} C \tau_p^2 \gamma + 4 \sqrt{Af}$$

$$F_\ell = E_{twist} + E_{bend} + E_{electro} = \frac{1}{2} C \tau_p^2 \ell + \frac{1}{2} A \frac{\sin^4 \alpha}{r^2} \ell + U \ell$$

$$F_\Gamma = E_{twist} + E_{bend} + E_{electro} = \frac{1}{2} C \tau_p^2 \Gamma + q_b \sqrt{Af} + U \Gamma$$

$$F_f = 4n\sqrt{Af} + q_D m \sqrt{Af}$$

# Free energy

$$L = L_s + n\gamma + m(\ell + \Gamma)$$

corrections to the work of the external force

$$F = F_s + n F_\gamma + m (F_\Gamma + F_\ell) + F_f - T S$$


---

entropic terms

$$U = U + \frac{1}{2} kT \left( \frac{kT}{Ar^2} \right)^{1/3}$$

confinement in a tube

$$F_\gamma = F_\gamma - \frac{1}{4} \text{Log} \left( \frac{4\pi^2 A (kT)^2}{d^4 f^3} \right)$$

loop size fluctuations  
(d=1nm)

$$S(L, \ell, n, m) = (n + m) \text{Log} \left( \frac{L - m\ell}{\gamma} - n - m \right) - \text{Log} n! - \text{Log} m!$$

Tonks hard core gas

## Free energy

$$F = F_s + n F_\gamma + m (F_\Gamma + F_\ell) + F_f - T S$$

study  $F$  under the two constraints:

$$L = L_s + n\gamma + m(\ell + \Gamma)$$

$$n = Lk = \frac{\tau_s}{2\pi} L_s + \frac{\tau_p}{2\pi} (m\ell + n\gamma + m\Gamma) + \frac{\sin 2\alpha}{4\pi r} m\ell + mp + n\Lambda$$


  
**Twist**    **Writhe**

we replace  $\tau_s, L_s$  to obtain :

$$F_{nm} = F_{nm} (\alpha, r, \tau_p, \ell)$$

## Computation method

$$m \neq 0 \quad F_{nm} = F_{nm}(\alpha, r, \tau_p, \ell)$$

$$m = 0 \quad F_{n0} = F_{n0}(\alpha, r, \tau_p)$$

reference state

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$$F_{00} = -gL + \frac{1}{2}C_s \left( \frac{2\pi L k}{L} \right)^2 L$$

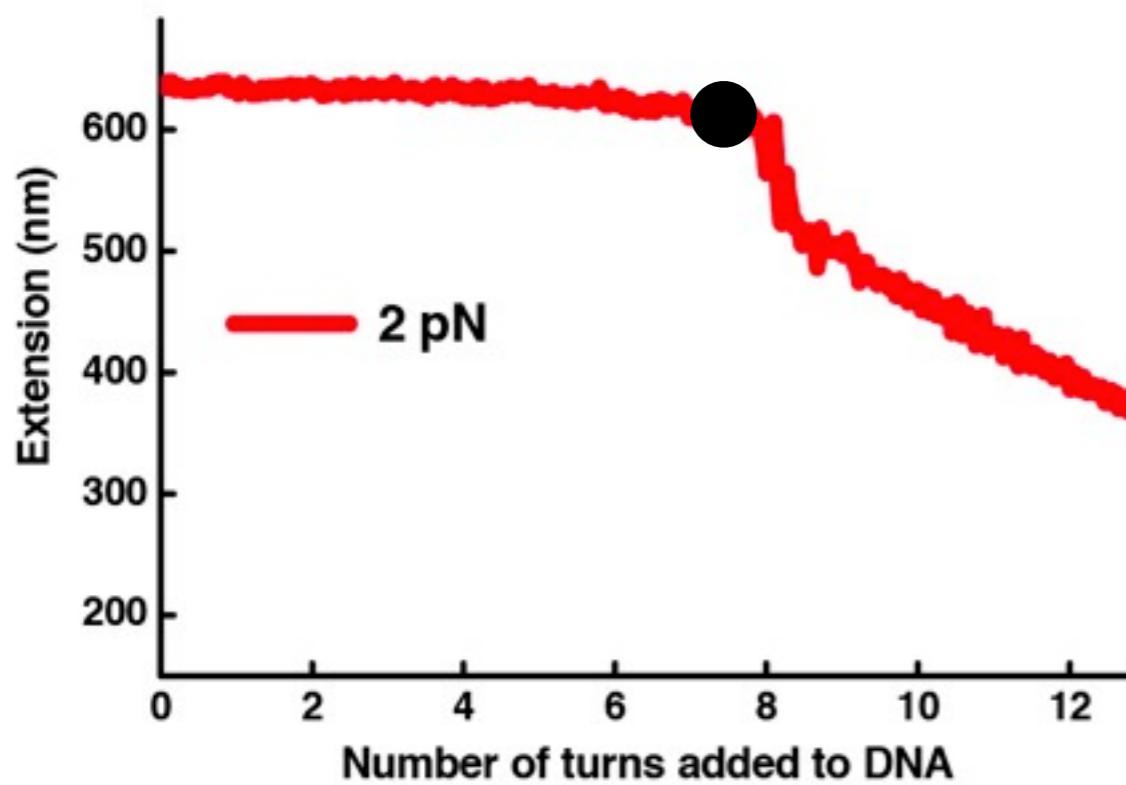
$$\Delta F_{nm}(\alpha, r, \tau_p, \ell) = F_{nm} - F_{00}$$

minimize with regard to  $\alpha, r, \tau_p$  (saddle point approx.)

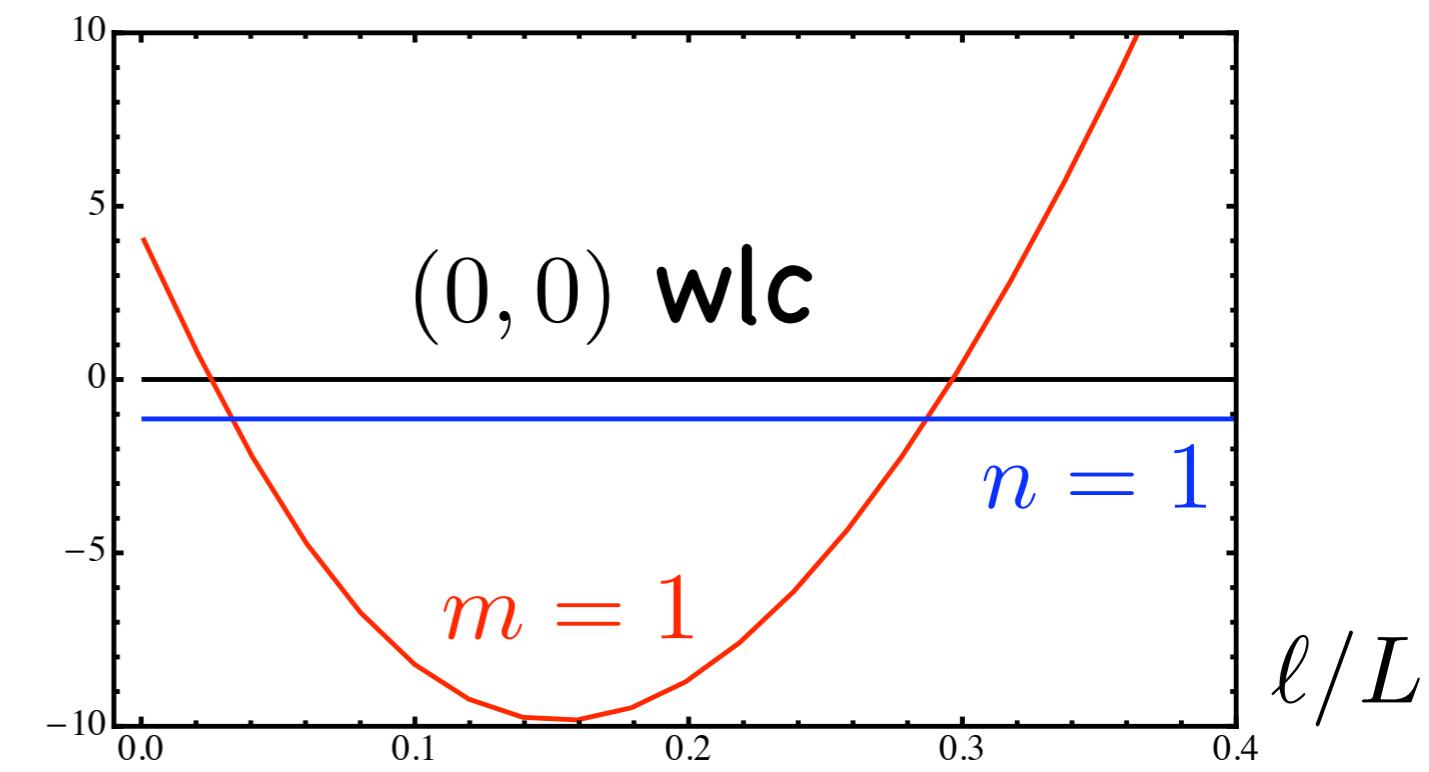
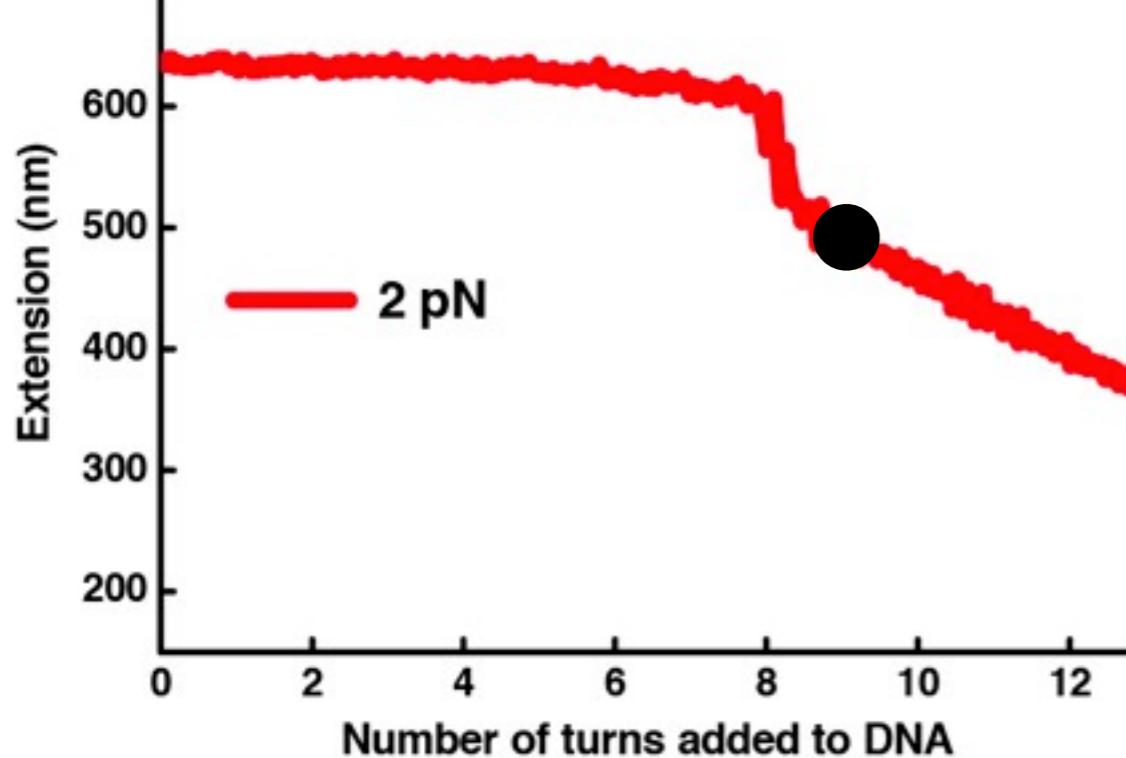
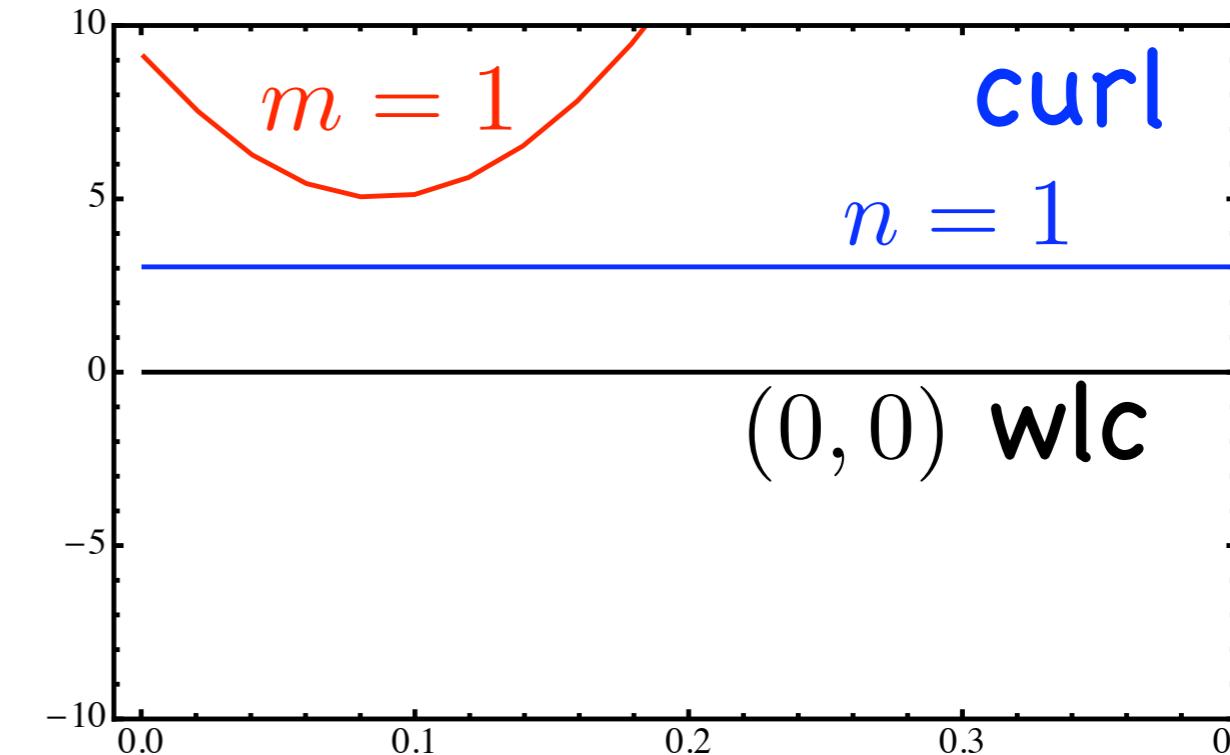
plot as function of  $\ell$

# Results

## Experimental data



$\Delta F/kT$  plecto



## Computing mean values

At fixed Lk

$$Z = \sum_n \sum_m \int_{\ell}^{\infty} e^{-F_{nm}(\ell)} d\ell$$

Probability of seeing a configuration with  
n curls and  
m plectonemic parts:

$$P_{nm}(n = Lk) = \frac{\int e^{-F_{nm}(\ell)} d\ell}{\sum_{n,m} \int e^{-F_{nm}(\ell)} d\ell}$$

Mean extension of the system (considering all  
possible configurations:

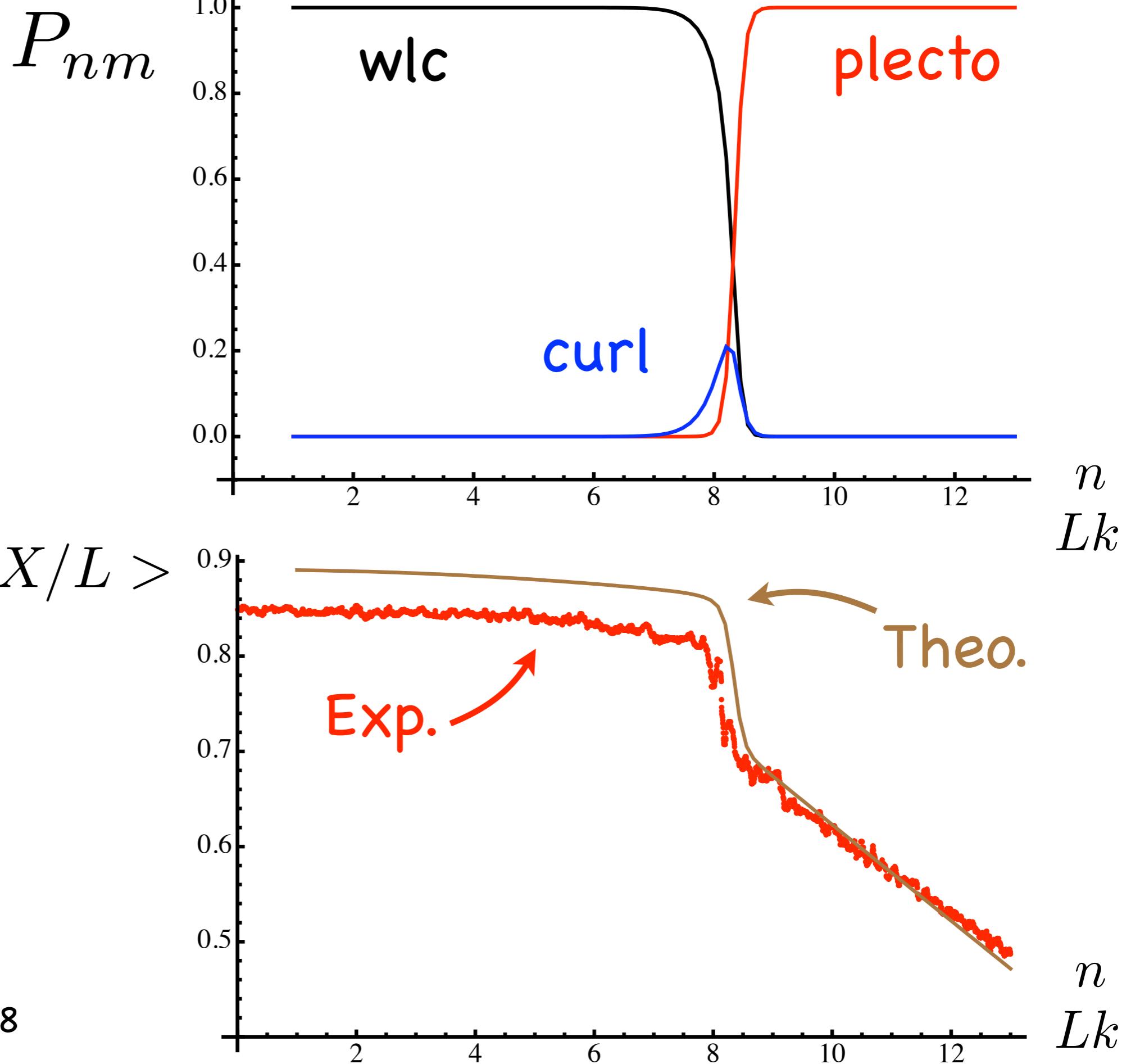
$$\langle X \rangle (n = Lk) = \frac{\sum_{n,m} \int X_{nm}(\ell) e^{-F_{nm}(\ell)} d\ell}{\sum_{n,m} \int e^{-F_{nm}(\ell)} d\ell}$$

# Results

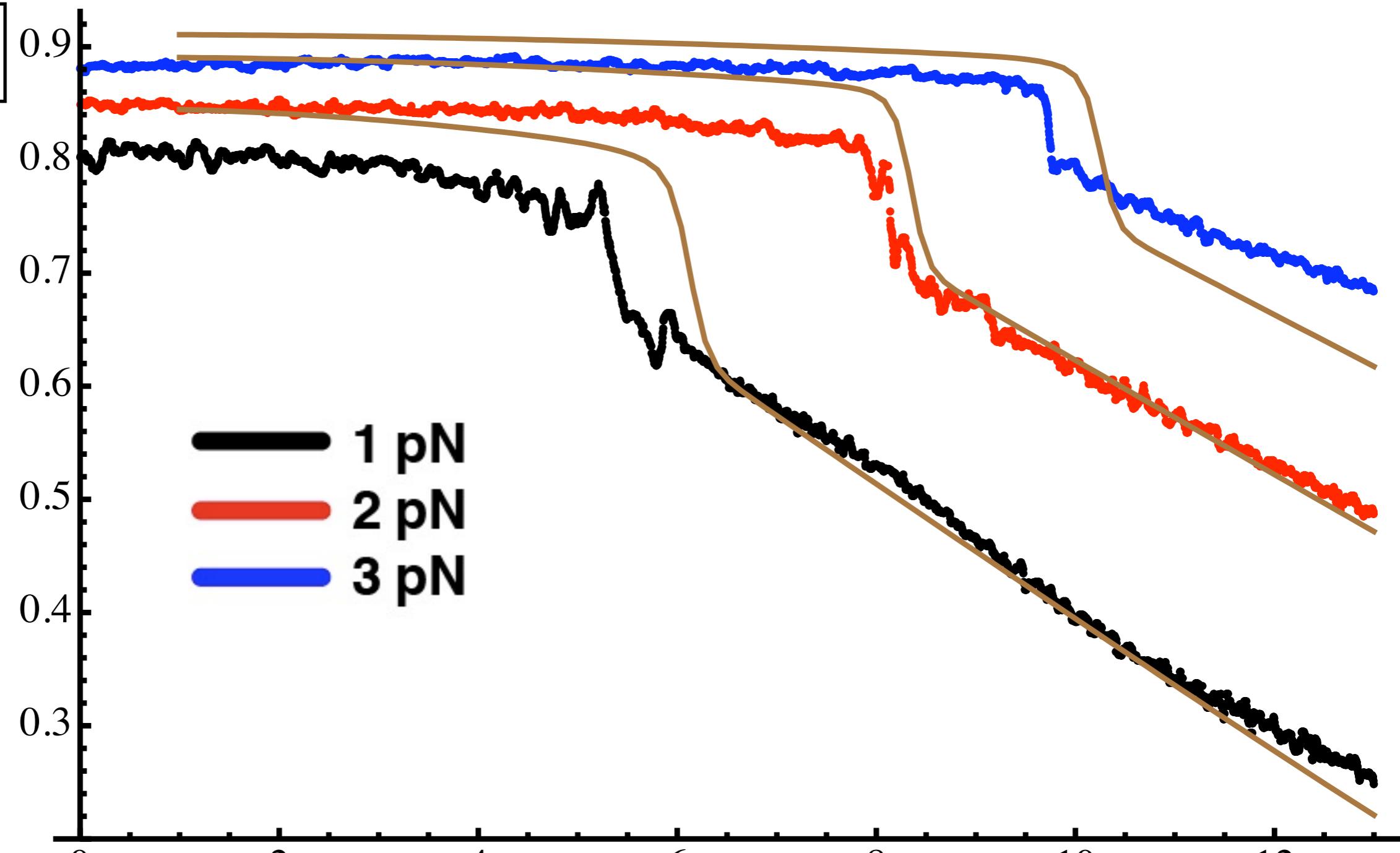
2.2 kbp  
150 mM  
2 pN

data : Forth et al  
(Phys. Rev. Lett.) 2008

$\langle X/L \rangle$



# Results

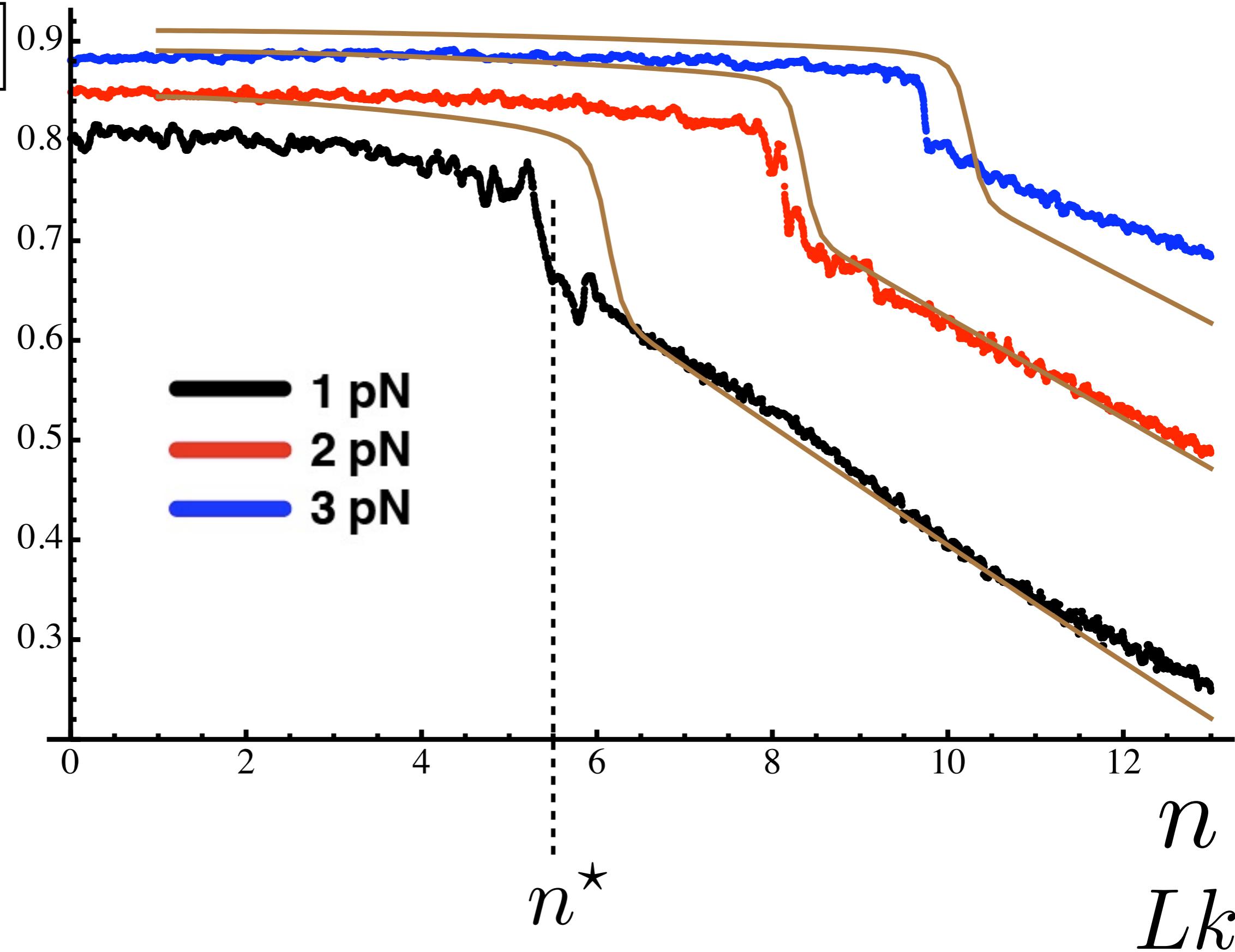


2.2 kbp  
150 mM

data : Forth et al  
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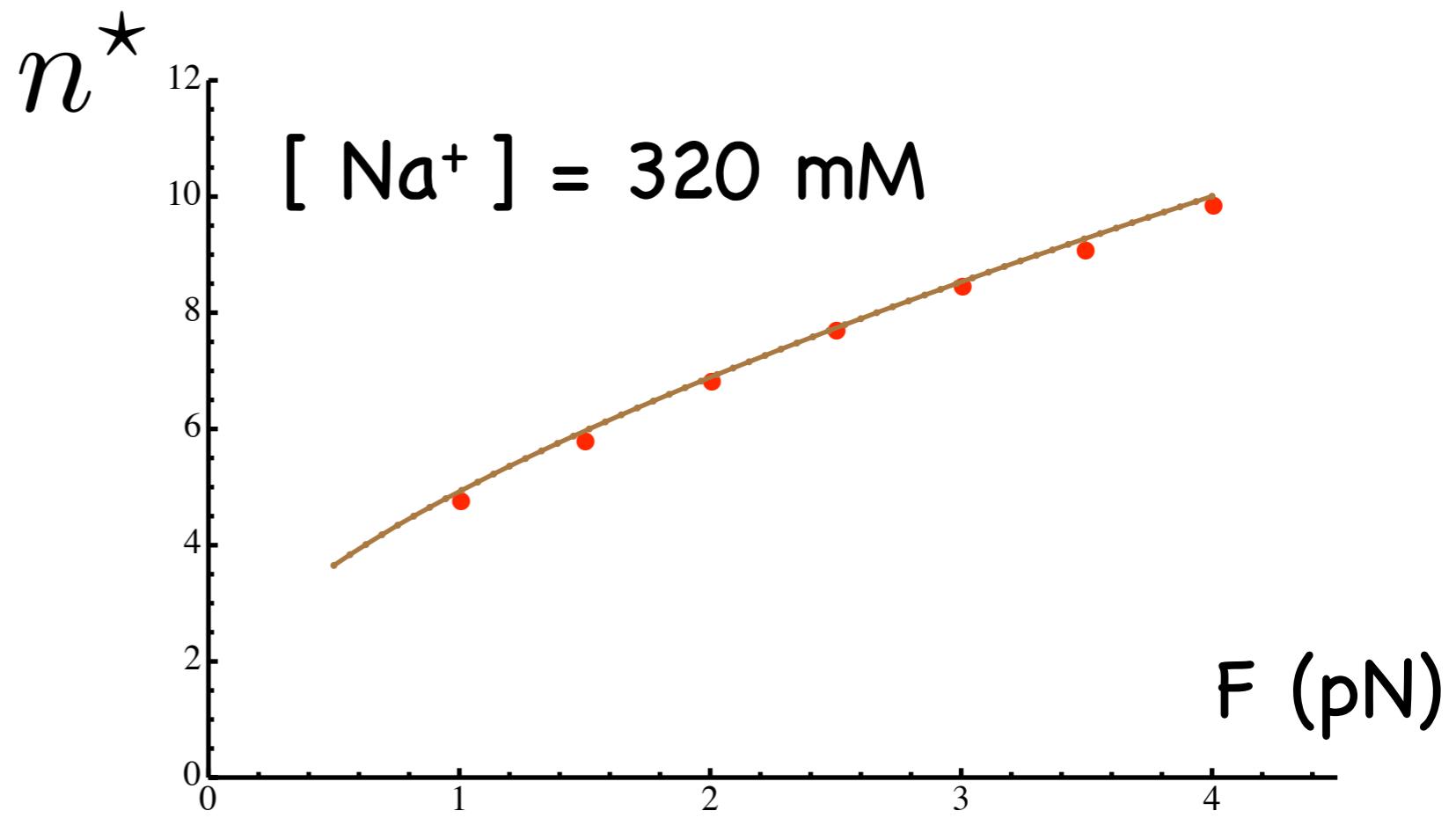
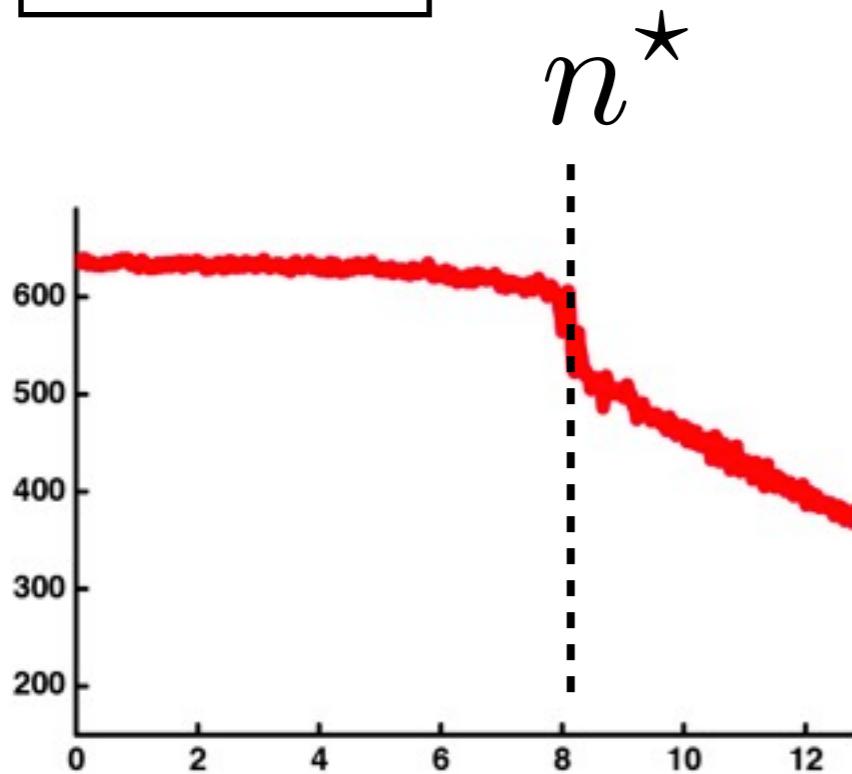
$n$   
 $Lk$

# Results



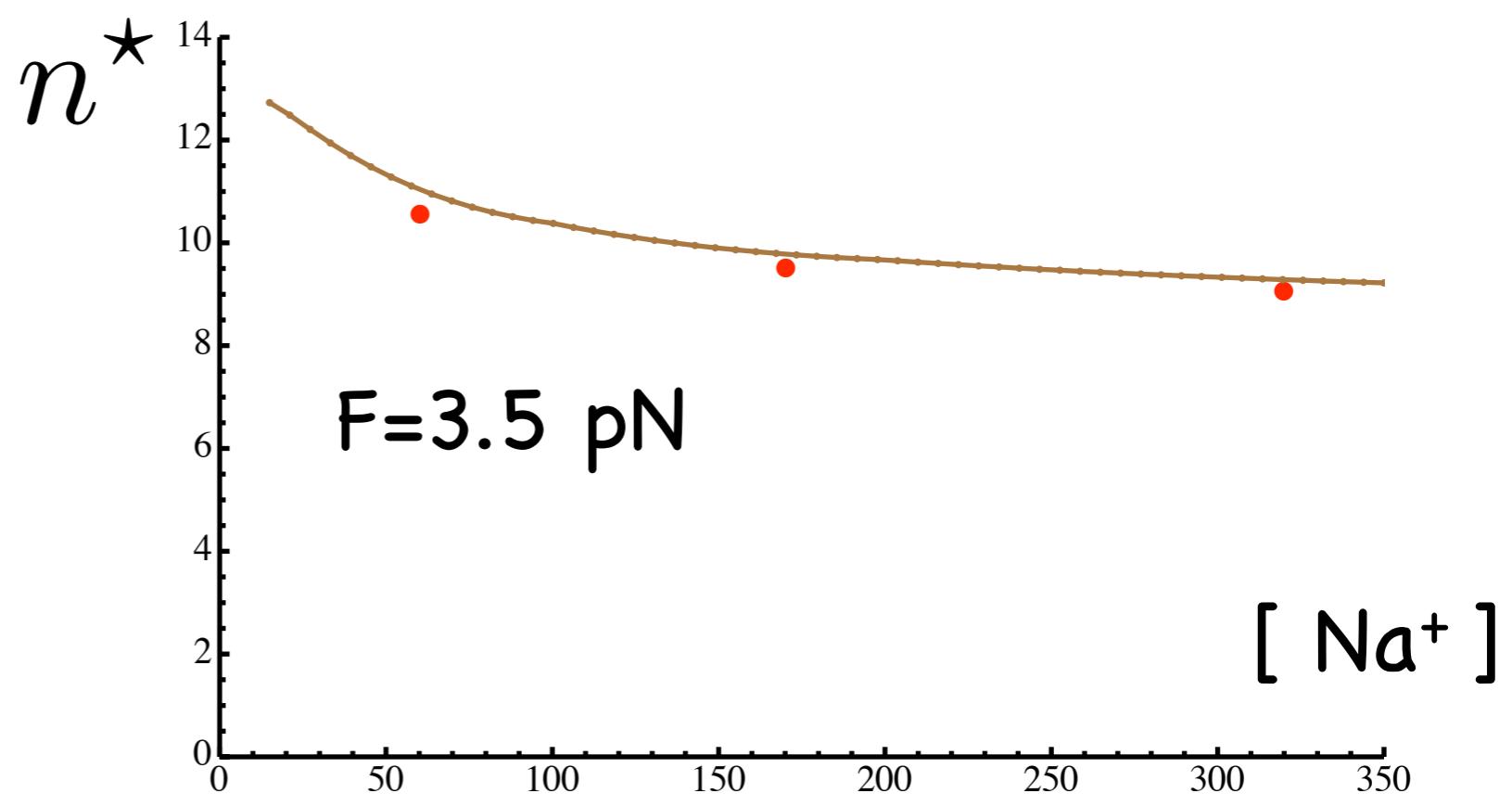
data : Forth et al  
(Phys. Rev. Lett.) 2008

# Results



1900 bp

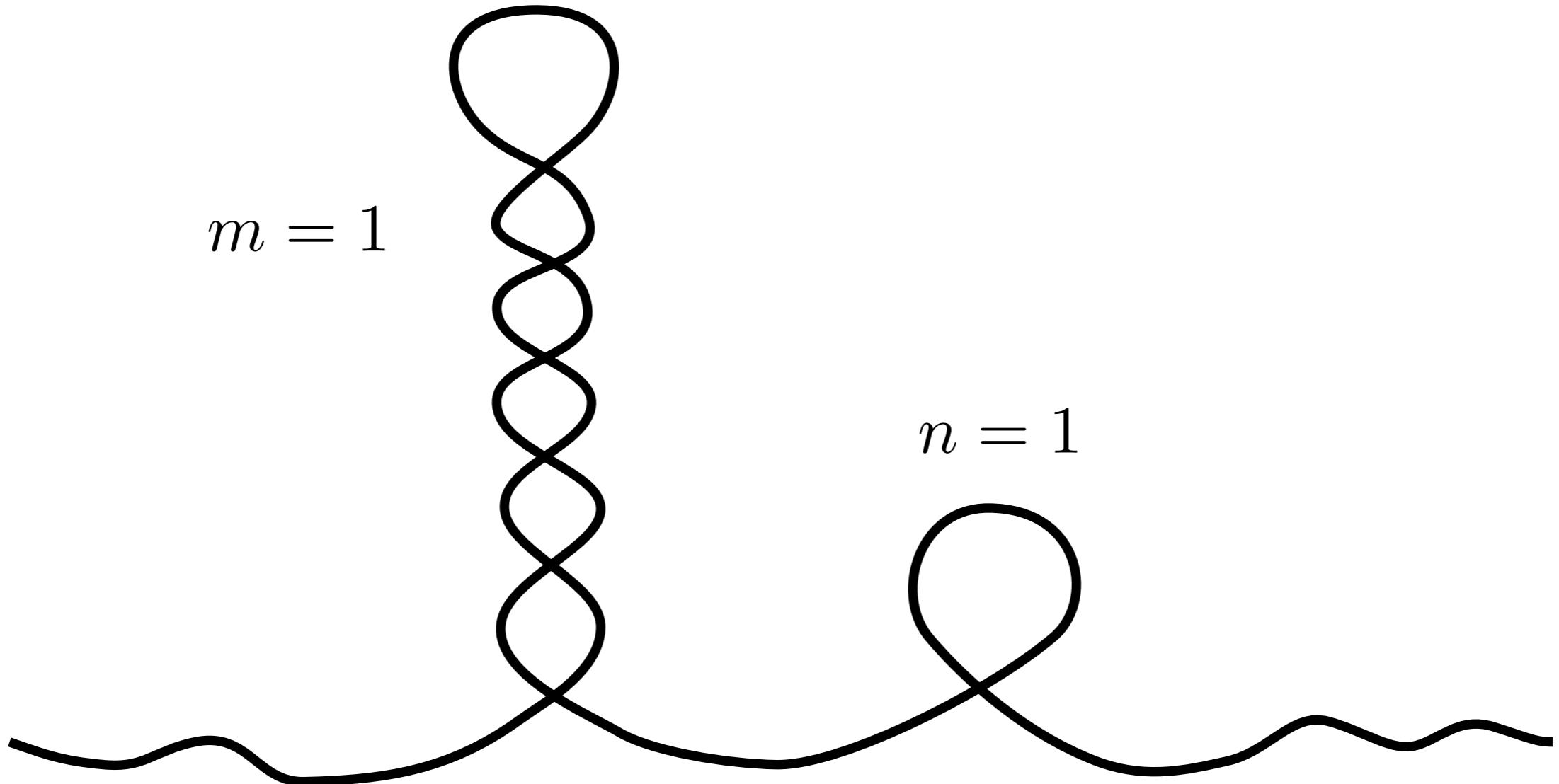
data : Brutzer et al  
(Biophys. J.) 2010



curls

Mean number of curls  
Mean number of plectonemes

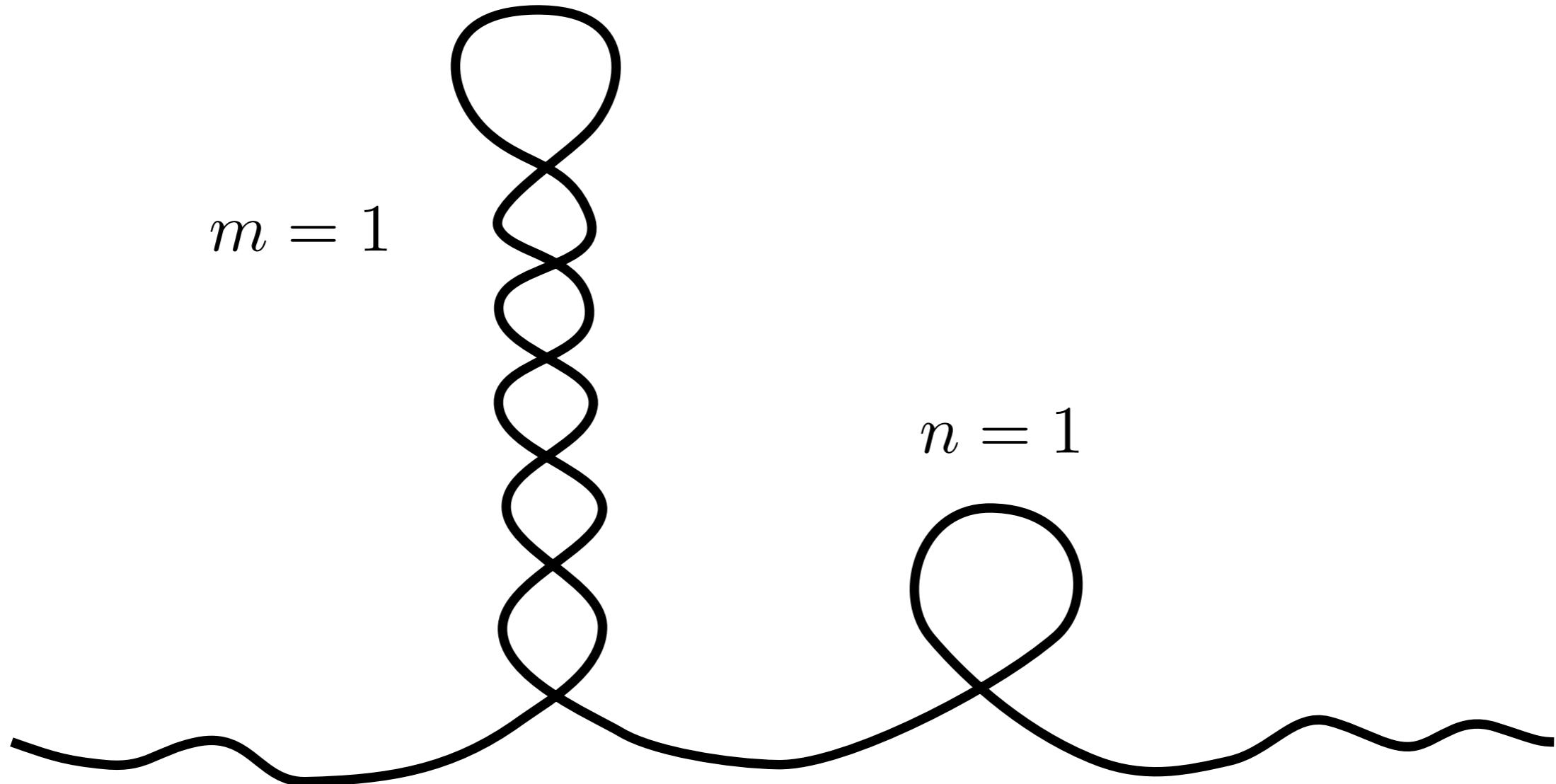
$$\langle n \rangle = \frac{\sum_{n,m} \int n e^{-F_{nm}(\ell)} d\ell}{Z}$$



Mean number of curls  
Mean number of plectonemes

plectonemes

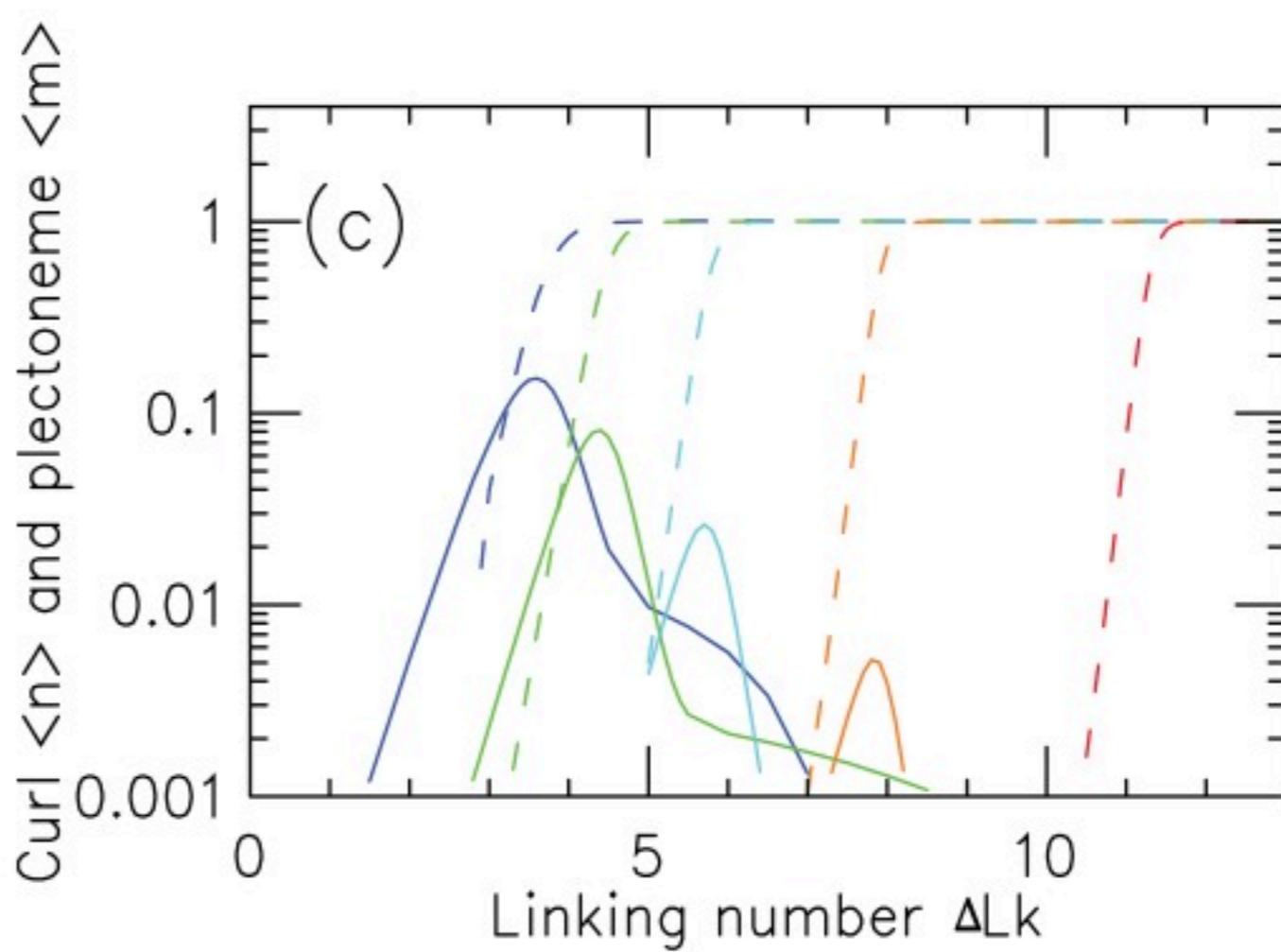
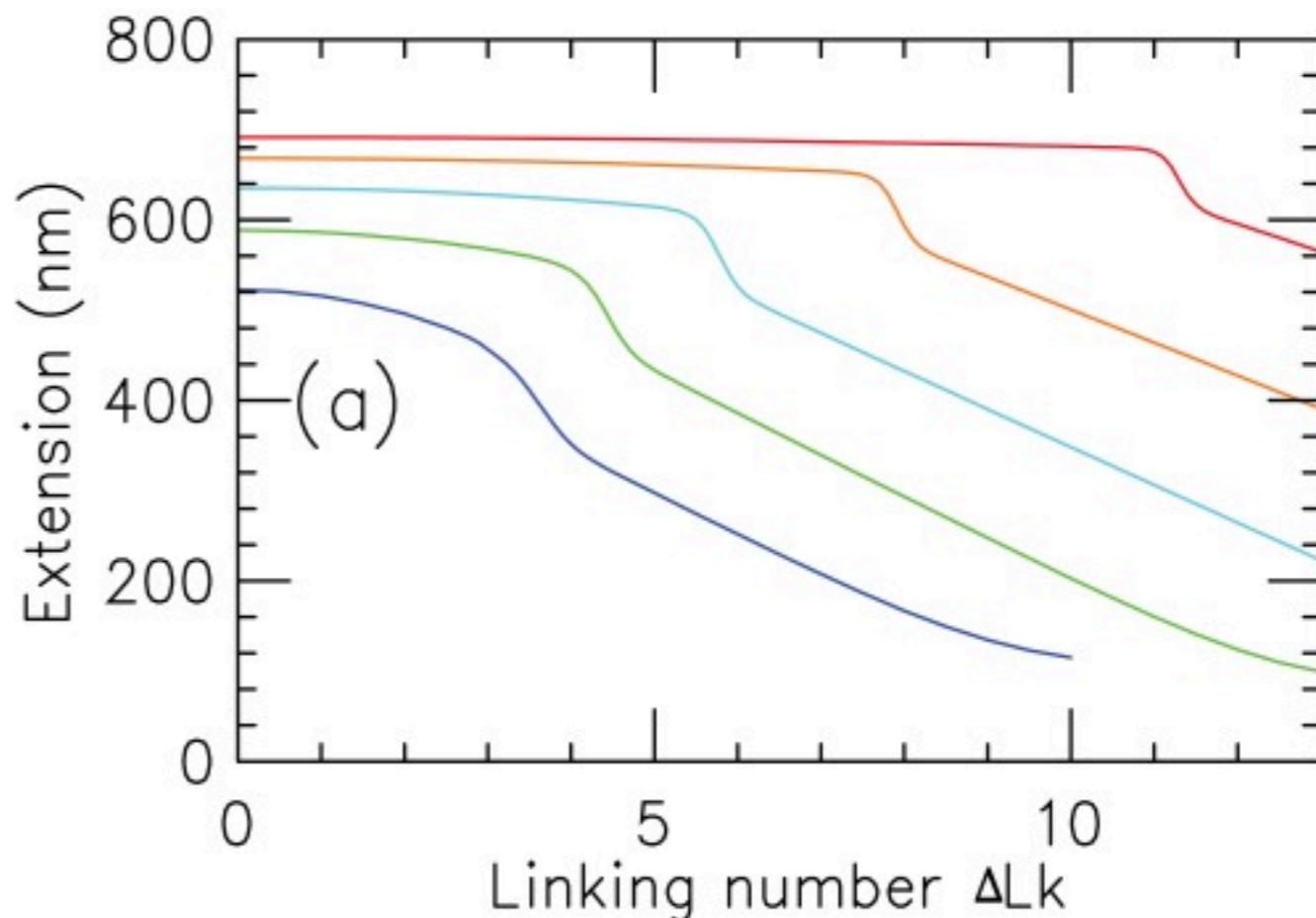
$$\langle m \rangle = \frac{\sum_{n,m} \int m e^{-F_{nm}(\ell)} d\ell}{Z}$$



2.2 kbp  
150 mM

0.25 pN blue  
0.5 pN green  
1 pN cyan  
2 pN orange  
4 pN red

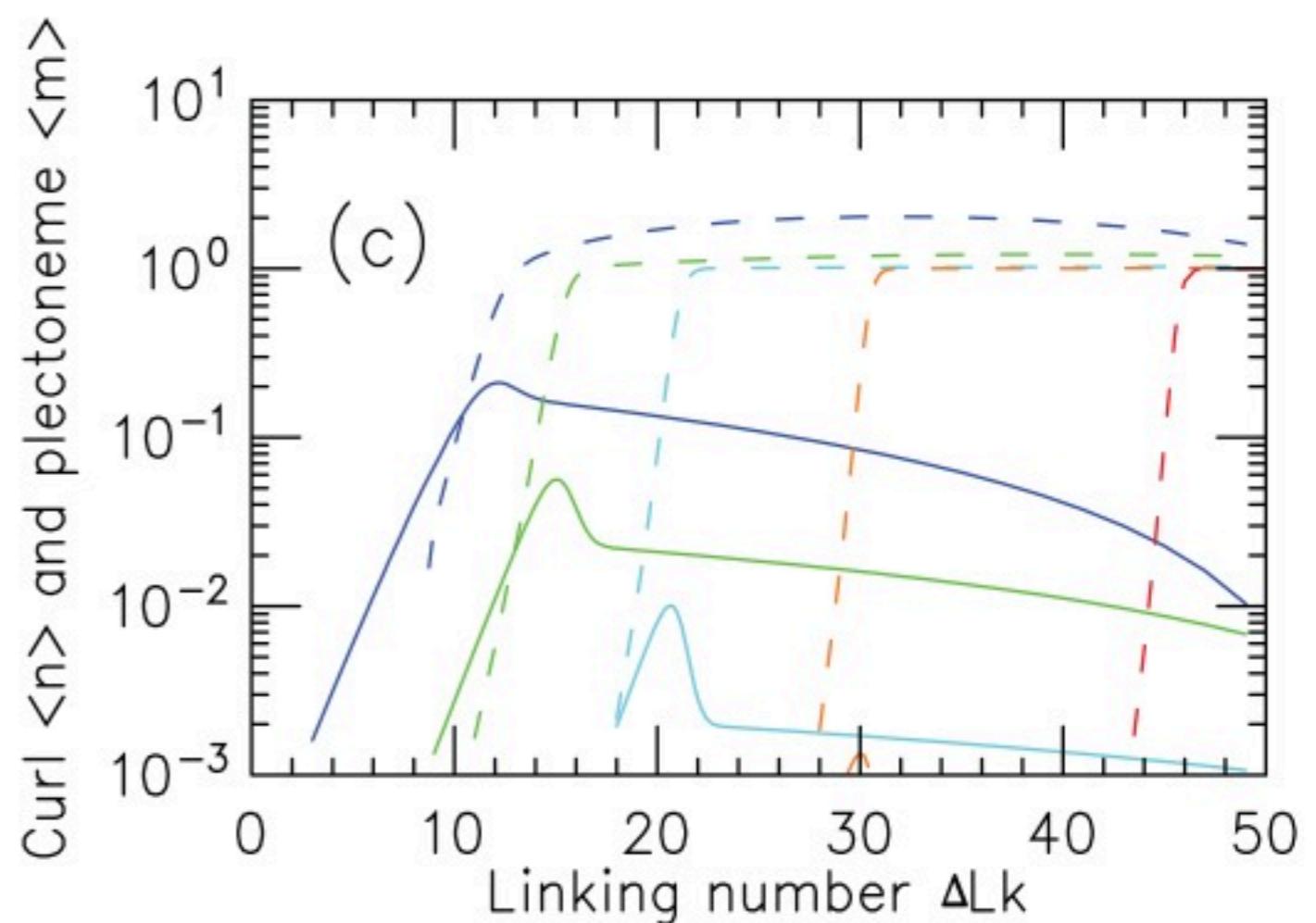
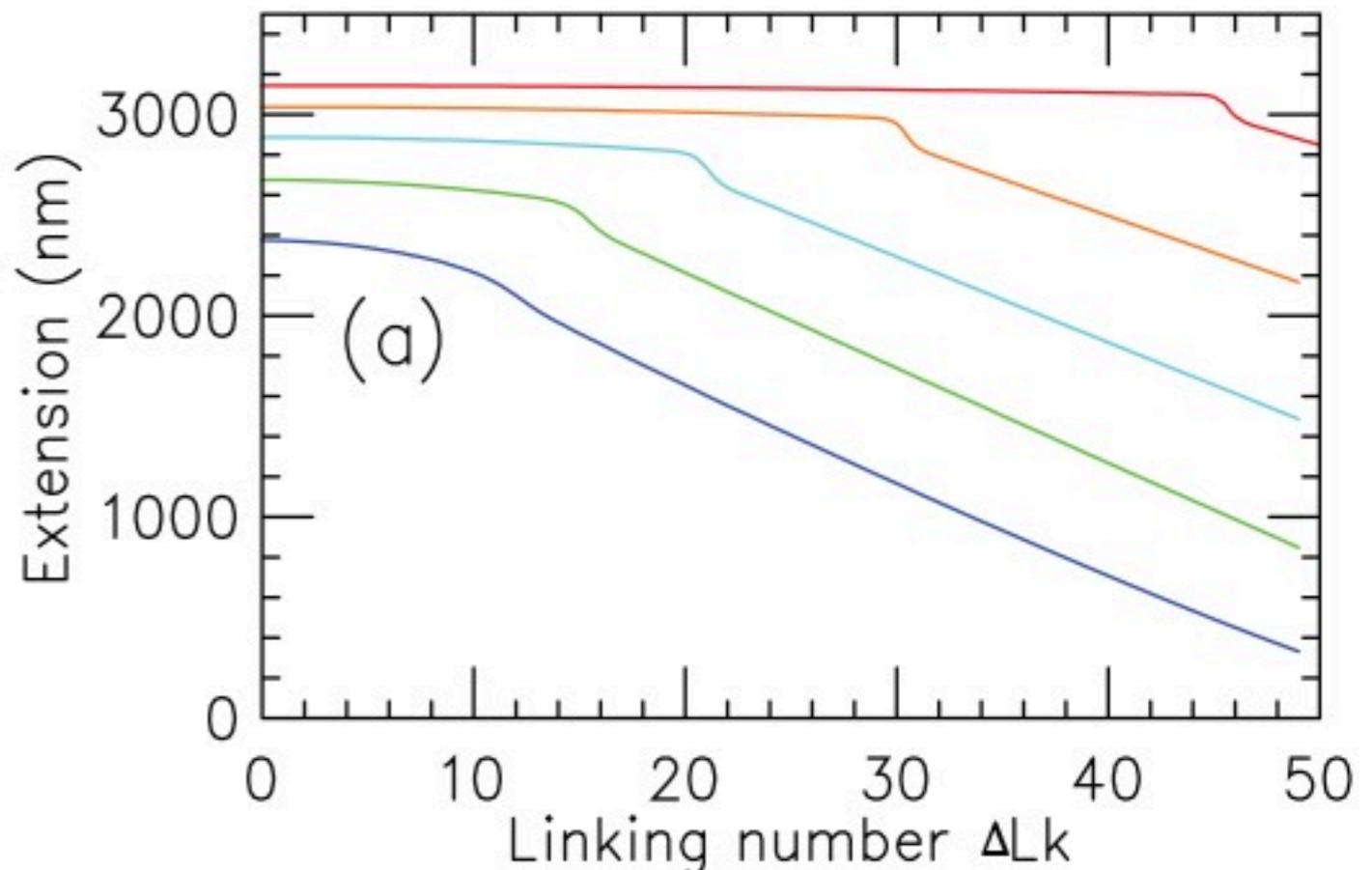
$\langle m \rangle$  dashed  
 $\langle n \rangle$  plain



10 kbp  
150 mM

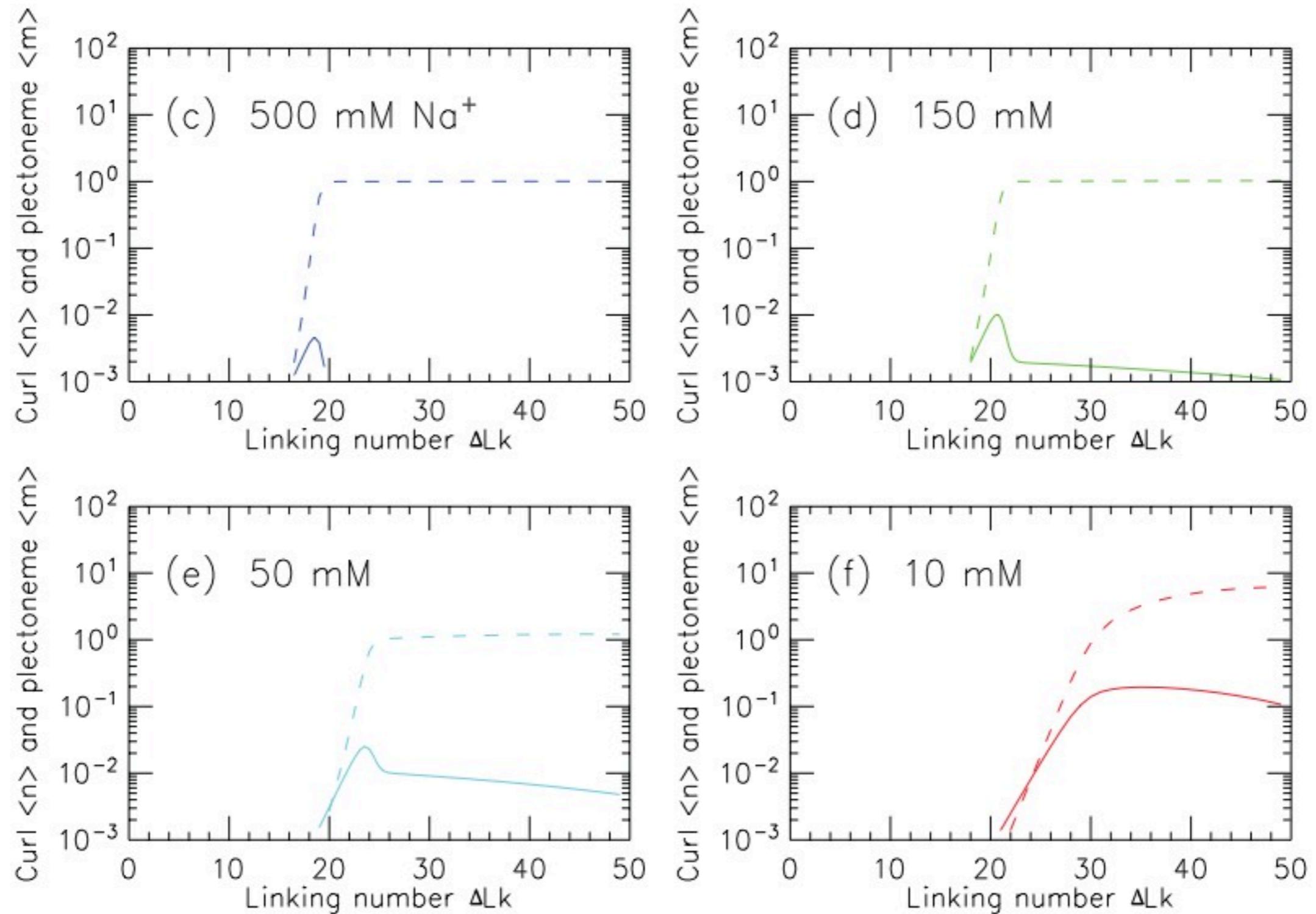
0.25 pN blue  
0.5 pN green  
1 pN cyan  
2 pN orange  
4 pN red

$\langle m \rangle$  dashed  
 $\langle n \rangle$  plain



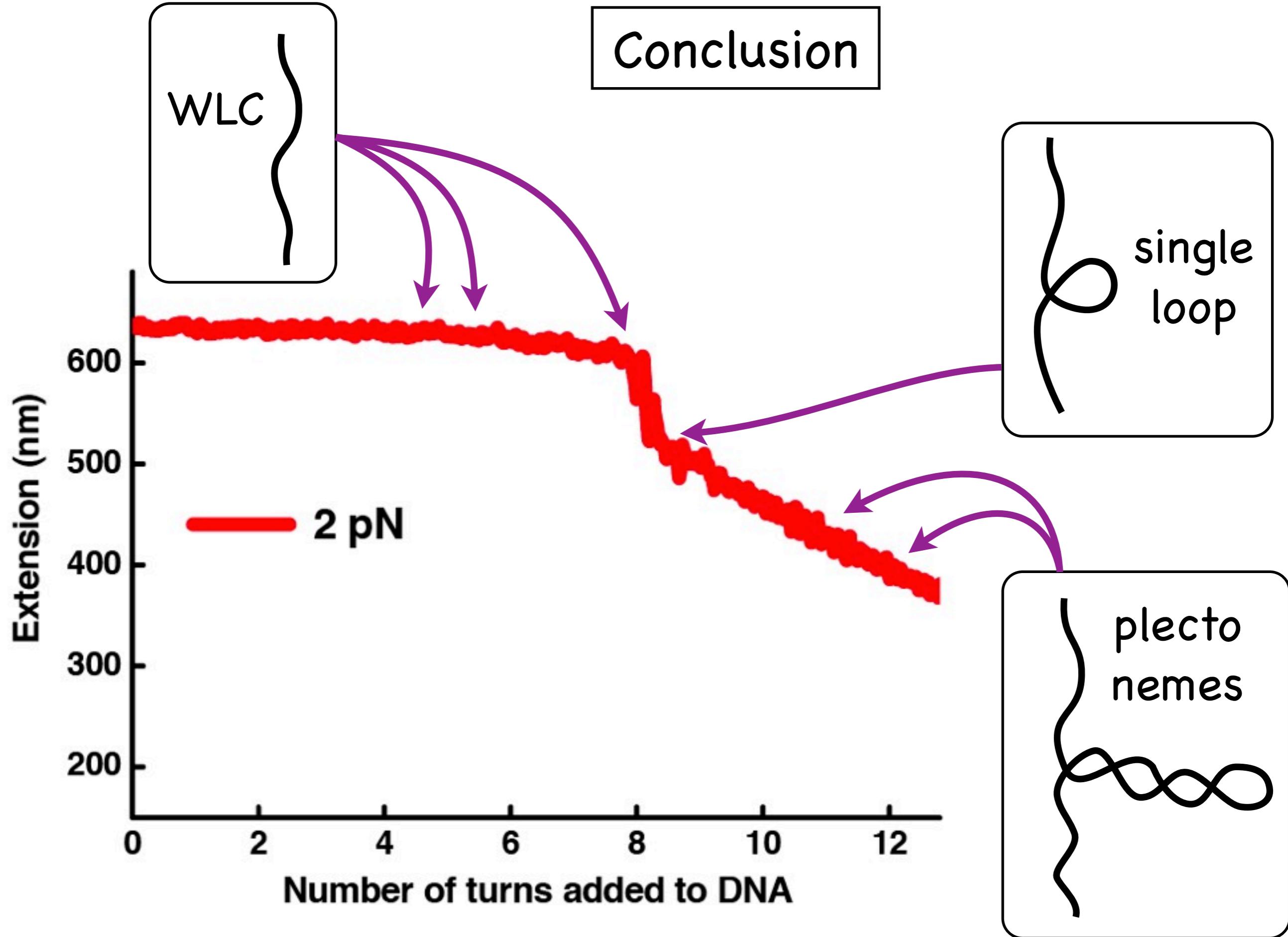
10kbp  
1 pN

# Salt dependence



=> lots of plectonemes and curls at low salt

## Conclusion



Thank you

Work was done with:  
N. Clauvelin (Rutgers)  
B. Audoly (Paris)  
J. Marko (Northwestern)